

Root Structural and Functional Dynamics in Terrestrial Biosphere Models – Evaluation and Recommendations

Journal:	<i>New Phytologist</i>
Manuscript ID:	NPH-TR-2014-17549.R2
Manuscript Type:	TR - Commissioned Material - Tansley Review
Date Submitted by the Author:	n/a
Complete List of Authors:	Warren, Jeffrey; Oak Ridge National Laboratory, Environmental Sciences Division and Climate Change Science Institute Hanson, P; Oak Ridge National Laboratory, Environmental Sciences Division and Climate Change Science Institute Iversen, Colleen; Oak Ridge National Laboratory, Environmental Sciences Division and Climate Change Science Institute Kumar, Jitendra; Oak Ridge National Laboratory, Environmental Sciences Division and Climate Change Science Institute Walker, Anthony; Oak Ridge National Laboratory, Environmental Sciences Division and Climate Change Science Institute Wullschleger, Stan; Oak Ridge National Laboratory, Environmental Sciences Division and Climate Change Science Institute
Key Words:	hydraulic redistribution, nutrient uptake, root function, root model, root plasticity, water uptake

Root Structural and Functional Dynamics in Terrestrial Biosphere Models – Evaluation and Recommendations

Warren, JM*, PJ Hanson, CM Iversen, J Kumar, AP Walker, SD Wullschleger

Environmental Sciences Division and Climate Change Science Institute, Oak Ridge National Laboratory, Oak Ridge, TN

* Corresponding author (Oak Ridge National Laboratory, P. O. Box 2008, Oak Ridge, TN 37831-6301, warrenjm@ornl.gov, 865-241-3150 telephone, 865-574-9701 fax)

Keywords: hydraulic redistribution, nitrogen uptake, root function, root model, root plasticity, water uptake

Running head: Modeling Root Function

Summary (201); Main Body (9271); Tables (3); Figs (5); References (204)

Notice: This manuscript has been authored by UT-Battelle, LLC, under Contract No. DE-AC05-00OR22725 with the U.S. Department of Energy. The United States Government retains and the publisher, by accepting the article for publication, acknowledges that the United States Government retains a non-exclusive, paid-up, irrevocable, world-wide license to publish or reproduce the published form of this manuscript, or allow others to do so, for United States Government purposes

Contents

Summary

I. Introduction

II. Current representation of root function in models

1. Single root models of water and nutrient uptake
2. Individual plant models of carbon allocation, architecture and resource acquisition
3. Ecosystem models
4. Terrestrial biosphere models

III. Recommendations for leveraging root knowledge into models

1. Scaling root function using root architecture
2. Linking root function to traits
3. Water uptake
4. Hydraulic redistribution
5. Ion uptake kinetics
6. Available root data – a serious limitation
7. Novel modeling platforms

IV. Conclusions

Acknowledgements

References

Summary

There is wide breadth of root function within ecosystems that should be considered when modeling the terrestrial biosphere. Root structure and function is closely associated with control of plant water and nutrient uptake from the soil, plant C assimilation, partitioning and release to the soils, and control of biogeochemical cycles through interactions within the rhizosphere. Root function is extremely dynamic and dependent on internal plant signals, root traits and morphology, and the physical, chemical and biotic soil environment. While plant roots have significant structural and functional plasticity to changing environmental conditions, their dynamics are noticeably absent from the land component of process-based Earth system models used to simulate global biogeochemical cycling. Their dynamic representation in large-scale models should improve model veracity. Here, we describe current root inclusion in models across scales, ranging from mechanistic processes of single roots to parameterized root processes operating at the landscape scale. With this foundation we discuss how existing and future root functional knowledge, new data compilation efforts, and novel modeling platforms can be leveraged to enhance root functionality in large-scale terrestrial biosphere models by improving parameterization within models, and introducing new components such as dynamic root distribution and root functional traits linked to resource extraction.

1 **I. Introduction**

2 Roots are key regulators of plant and ecosystem function through their role in water and nutrient
3 extraction from soils, and through the plasticity of their responses to changing resource
4 availability or environmental conditions (Hodge 2004, Schenk 2005). In this capacity, roots act
5 as a key mediator of vegetation evapotranspiration, which dominates the control of land surface
6 energy and water balances. Similarly, through uptake of nitrogen and other nutrients, roots are
7 critical for biogeochemical cycling and the interwoven carbon cycle that regulates C balance
8 (Fig. 1). Our knowledge of root functional processes is extensive and continues to improve with
9 new research initiatives and advanced experimental techniques.

10
11 Notwithstanding the many important roles of roots, dynamic root functions are still largely
12 absent in land surface models (Woodward and Osborne 2000, Ostle et al. 2009, Matamala and
13 Stover 2013, Iversen 2014), hereafter referred to by the more inclusive term Terrestrial Biosphere
14 Models (TBMs). Root representation in TBMs is rudimentary, with carbon allocation, root
15 distribution, water uptake and nutrient (almost solely limited to nitrogen) extraction generally
16 based on fixed parameters or plant demand, independent of dynamic root functionality. Key root
17 attributes that are missing include the capacity of roots to shift distribution under changing
18 environmental conditions, regulate water uptake (e.g., via aquaporins), regulate nutrient uptake
19 (e.g., via enzyme-mediated Michaelis-Menten kinetics), or associate with mycorrhizal fungi. The
20 limited representation of roots in TBMs is partially due to a lack of appropriate global root
21 datasets, but also due to the fact that TBM representation of vegetation processes under current
22 climatic conditions appears to work fairly well with little or no representation of roots. TBMs use
23 of implicit parameters of bulk water and nutrient uptake independent of roots can correlate to
24 total root uptake (Norby and Jackson 2000, Woodward and Osborne 2000, Feddes et al. 2001),
25 and requires minimal root data or computational resources. Yet while the simplified models may
26 be roughly adequate, they do not allow dynamic root functionality, and thereby (we believe)
27 limit application to future environments, and limit mechanistic linkages that establish model
28 validity. Without inclusion of root dynamics, the current representation of roots in TBMs may

29 not be sufficient to capture their roles in ecosystem function, nor adequate to understand
30 potential controls that expressed root function may have in response to environmental change.

31
32 Feddes et al. (2001) argued “that the functioning of roots [...] needs to receive more attention in
33 land surface and climate modeling.” Model representation of canopy structure and function has
34 progressed significantly (e.g. Mercado et al., 2007; Bonan et al., 2011; Loew et al., 2013) since
35 the big-leaf approach cited by Feddes et al. (2001). Alternately, and with some notable
36 exceptions (e.g., hydraulic redistribution of water, Lee et al. 2005; multi-process nitrogen uptake,
37 Fisher et al. 2010), the representation of root structure and function in TBMs has seen only
38 limited progress. Improved representation of root water uptake has stalled despite demonstration
39 of model sensitivity to roots in climate and vegetation distribution simulations over a decade ago
40 (Kleidon & Heimann, 1998, 2000; Hallgren & Pitman, 2000, Feddes et al. 2001).

41
42 In contrast to simplified TBM’s that must represent the dynamics of roots associated with the
43 entirety of the global land surface, mechanistic models at the scale of single root processes
44 include the necessary complexity to capture water and nutrient uptake functions in response to
45 environmental stimuli at quite high resolution in both space and time (Gardner 1960, Barber
46 1962, Hillel et al. 1975, Raats 2007). Higher-order model development often makes simplifying
47 assumptions about such processes, potentially missing a fundamental control point for plant
48 function under varying resource availability.

49
50 A whole universe of knowledge on root characteristics and functions exists that has not been
51 exercised within TBMs. Novel nondestructive imaging techniques of roots have provided new
52 insights in form and function of roots *in situ* (Fig. 2). Confocal laser microscopy has been used to
53 assess dynamic gene expression of root initiation and cell growth within the root tissues (Busch
54 et al. 2012, Vermeer et al. 2014). Linked studies of gene regulation, growth regulators,
55 intercellular communication and tissue development have led to advances in mechanistic
56 multiscale modeling that can be used to predict root phenotypes (Band et al. 2012). Actively
57 controlled root membrane aquaporins have been identified as implicit control points for water

58 transfer across roots (Javot and Maurel 2002; Maurel et al. 2008). Next generation
59 minirhizotrons are yielding unprecedented insight into fine-root and mycorrhizal exploration and
60 turnover at high temporal resolution (Allen and Kitajima 2013), and have been paired with CO₂
61 sensors to allow concurrent measurements of respiration in situ (Vargas and Allen 2008).
62 Neutron imaging has recently been used to assess in situ soil-root-rhizosphere hydration
63 (Carminati et al. 2010) and individual root water uptake and transport dynamics (Warren et al.
64 2013). Soil moisture sensors continue to evolve, and allow for highly precise measurements of
65 root water extraction dynamics and hydraulic redistribution throughout the soil profile (e.g.,
66 Warren et al. 2011). Such measurements provide insight into soil, rhizosphere and root
67 resistances, data that can be used to refine models of physical flow of water through the soil-
68 plant system (Gardner 1965, Sperry et al. 1998). Other root functional processes including C
69 flux, water and ion uptake, water potential and rhizosphere nutrient competition have been
70 elucidated using novel biosensors (Herron et al. 2010), isotope tracers (Bingham et al. 2000), and
71 *in situ* field observations (Lucash et al. 2007). Despite this extensive knowledge of single root
72 processes, the scaling of such processes spatially within the soil profile and across the landscape
73 through time has not been achieved.

74
75 The knowledge gap that exists in mechanistic model representation of root processes across
76 scales (i.e., between roots, individual plants, ecosystems or land surfaces) is in part a
77 consequence of inadequate datasets and the difficulty in linking root function to characteristic
78 root traits, root distribution and root growth dynamics across landscapes (Fig. 3). For model
79 veracity, simplified processes modeled in TBMs "...should be based on mechanistic
80 understanding of the processes at lower scales..." (Schulze 2013) – an understanding that has not
81 been well-translated for roots or root function. As such, the gap in knowledge transfer across
82 scales leads to decreases in the expression of detailed root function as the predictive scale of the
83 model increases (Ostle et al., 2009). To model climate and the Earth System, TBMs must
84 simulate the land surface energy, water and carbon balances at broad spatial (e.g., km)
85 resolutions and at timescales ranging from every 15 minutes to potentially several hundred years
86 (Pitman, 2003). The models must therefore integrate across the microscopic (e.g., sub-mm) and

87 comparably short-term (e.g., seconds to minutes) scales relevant for actual root tissue function.
88 Thus the *microscopic*, mechanistic approach of single root modeling is not readily scaled to the
89 landscape, which led to development of *macroscopic*, bulk, sink-based modeling (Skaggs et al.,
90 2006) at the plant or ecosystem scales.

91
92 This review considers how root function is represented by models across scales, ranging from
93 single roots to whole land surfaces, and provides recommendations for improved representation
94 of roots in TBMs. The current state of knowledge regarding root structure and function is
95 considered, and the inherent and dynamic plasticity in those characteristics is described.
96 Leveraging this mechanistic knowledge, a focus was placed on identifying aspects of root
97 structure and function that could affect root water and nutrient uptake dynamics in context of
98 carbon cycling within TBMs. Specific targets for model improvement are noted. Since data are
99 required for model parameterization and validation, data availability is examined as a limitation
100 of the application of root function into models across scales. The scope of the review was limited
101 to living root characteristics that directly affect whole plant function, including growth, and ion
102 and water uptake. The indirect implications of root exudation, turnover and rhizosphere ecology
103 (Young 1998, Cheng et al. 2014), while critically important, were not considered in this review.

104

105 **II. Current representation of root function in models**

106 1. Single root models of water and nutrient uptake

107

108 Single root water uptake occurs across a diversity of spatial scales requiring different approaches
109 to best model water extraction. The microscopic approach involves physical first-principle
110 mechanistic descriptions of radial flow to, and uptake by, individual roots (Hillel et al., 1975). In
111 contrast, the macroscopic approach models uptake with a sink term in the Richards equation that
112 ignores or implicitly averages uptake over a large number of roots (Skaggs et al., 2006). Early
113 experimental and modeling work was carried out by Gardner (1960) where a root was modeled
114 to be an infinitely long cylinder of uniform radius and water uptake characteristics. Although this
115 formulation of root water uptake stimulated much research (Gardner, 1964 and 1965), it was

116 soon emphasized that it was not practical to develop field-scale models of water transport if flow
117 to each individual root of a complete root system must be considered (Molz and Remson, 1970;
118 Molz, 1981). Thus, various extraction term models have been developed where the fundamental
119 premise is to describe root water uptake for the rooting zone rather than individual roots. In these
120 models, soil-root processes are generally reduced to a root sink term that is incorporated into a
121 detailed description of soil water balance (Doussan et al., 2006).

122
123 Classical models of nutrient acquisition at the scale of a single root have provided many insights
124 into the complex dynamics that occur at the root-soil interface. Early pioneering research by
125 Barber (1962), Nye (1966), and Nye and Marriott (1969) indicated that nutrient uptake could be
126 modeled as a single cylindrical root in an infinite extent of soil, where diffusion and mass flow
127 supply nutrients to the root absorbing surface (Rengel, 1993). In most models that derive from
128 the Nye-Barber framework, the central hypothesis is that the driving force of nutrient acquisition
129 is the absorption of nutrients by the root, which results in a decrease in nutrient concentration at
130 the surface of the root, leading to a diffusion gradient and movement of nutrients in the soil pore
131 water (Hinsinger et al., 2011). Although early models were confirmed by kinetic studies using
132 plants grown in hydroponic culture, the difference between nutrient acquisition between well-
133 stirred solution and heterogeneous soil are large (Rengel, 1993). As a result, uptake can be
134 overestimated by these models because nutrient concentrations calculated at the root surface may
135 be too high.

136
137 While the pioneering studies of single-root water and nutrient uptake established the modeling
138 framework for basic root resource acquisition, a wealth of new knowledge from genomic to
139 cellular to whole root scales has emerged over the last several decades and improved our
140 understanding of root structure and function (Fig. 1, 2). These insights offer novel understanding
141 of single root functional plasticity that might be leveraged into better representation in TBMs (as
142 discussed later).

143

144 2. Individual plant models of carbon allocation, architecture and resource acquisition

145 Whole-plant models require more sophisticated approaches and involve a higher level of
146 complexity in the description of root structure and function than single root models. These
147 approaches include an expanded consideration of how photosynthate is allocated to roots given
148 competing sinks, and how the processes of root tip initiation, branching, and geotropism give rise
149 to three-dimensional patterns of root distribution in soils (e.g., Thaler and Pagès, 1998, Ge et al.
150 2000).

151
152 Various models have been developed over the last 25 years to describe the structure and function
153 of whole plant root systems (Clausnitzer and Hopmans, 1994; Jourdan and Rey, 1997; Spek,
154 1997; Dupuy et al., 2007; Dupuy et al. 2009, Schnepf et al., 2012). Five models in particular
155 stand out as addressing the comprehensive suite of processes that govern photosynthate
156 allocation to root growth, root system architecture, and acquisition of water and nutrients from
157 heterogeneous soils (Table 1). These models simulate the production of daily photosynthate and
158 its allocation to plant organs based on general source-sink concepts (Franklin et al., 2012).
159 Growth and respiration of leaves, stems, and roots are often represented as competing sinks for
160 photosynthate. The SPACSYS model (Wu et al., 2007) is an exception in that roots receive
161 photosynthate with the highest priority, followed by leaves then by stems. Interestingly, several
162 models include options for allocation of photosynthate (Table 1). Most notable is the scheme
163 implemented in Root Typ (Thaler and Pagès, 1998), where allocation can be modeled either as a
164 function of competing sinks (i.e., without priorities) or where photosynthate is totally allocated to
165 meet the demands of all plant organs. Each of the root growth models described in Table 1 can
166 provide realistic spatial complexity of root system architectures consisting of distinct root
167 classes (Wu et al., 2007; Pagès et al., 2004; Postma and Lynch, 2011a), where each root is
168 represented by a growing number of root segments interacting with the soil. Comparison of
169 model results with visual images from excavated plants (Clausnitzer and Hopmans, 1994; Pagès
170 et al., 2004; Wu et al., 2007) and measured root density by depth (Somma et al., 1998) provide
171 encouraging support for the realism and utility of these simulations.

172

173 The ability to model root architecture allows coupling of root distribution with mechanistic
174 descriptions of water and nutrient uptake (Table 1) (Dunbabin et al., 2004; Ho et al., 2004; Janott
175 et al., 2011). For example, the R-SWMS model has been used to simulate the dynamic and
176 spatial patterns of root water extraction (Draye et al. 2010). Results indicated that it was the
177 interplay between root architecture, root axial and radial hydraulic properties, and water
178 distribution in spatially heterogeneous soils that controlled patterns of water extraction. The
179 *SimRoot* model has been coupled to a phosphorus acquisition and inter-root competition model
180 (Ge et al. 2000). Results indicated that phosphorus acquisition differed across different root
181 system geometries, with greater phosphorus uptake per unit carbon cost for shallow root systems
182 compared to deeper root systems. In similar fashion using ROOTMAP, Dunbabin et al. (2003)
183 found that the optimal root architecture for nitrate capture in sandy soils was one that quickly
184 produces a high density of roots in upper soils to facilitate nitrate uptake during the early season,
185 but also had a vigorous taproot growth for nitrate acquisition later in the season.

186
187 Two or three-dimensional modeled root architecture frameworks could be further refined to
188 allow differential plasticity in growth and function that might be incorporated into future models,
189 especially if dynamic root water and nutrient uptake capacity could be assigned based on root
190 age, root order, or differential hydraulic conductivity (Valenzuela-Estrada et al. 2008). Indeed,
191 two-dimensional bulk soil water uptake has been successfully modeled as a series of resistances
192 through the soil, root, plant and atmosphere continuum, regulated by water potential gradients
193 and verified with field data (Sperry et al. 1998, Hacke et al. 2000, Wang et al. 2002, Manzoni et
194 al. 2013). Manoli et al. (2014) introduced a three-dimensional model based on pathway
195 resistances that includes hydraulic redistribution and that allows root systems of multiple trees to
196 compete for water extraction from different soil layers. Such models are noteworthy in that they
197 retain first principle, physics-based *Darcian* water flow at the stand level, while allowing
198 dynamic root functionality under drying conditions, a feature often lost in ecosystem models.

199

200 3. Ecosystem models

201 While root and individual plant models are highly-detailed, they usually do not have the
202 appropriate temporal and spatial resolution to simulate plant interactions with the surrounding
203 soil at the ecosystem level (Agren et al., 1991). Ecosystem process models were developed to
204 simulate feedbacks and linkages among ecosystem components (plants, microbes, and resource
205 pools) to assess whole ecosystem C, water, and nutrient cycling across biomes such as forest
206 stands (Running & Coughlan, 1988) or grasslands (Parton et al., 1988). While ecosystem process
207 models encompass spatial scales and processes ranging from the plot level (Running &
208 Coughlan, 1988) to the global land surface (Hopkins and Bristow, 2002), they are distinct from
209 TBMs in that they are not generally intended to be scaled to the global land surface or informed
210 with products of remote sensing (Running & Coughlan, 1988). However, many ecosystem
211 process models were developed to interface with TBMs (Parton et al., 1988; Riley et al., 2009;
212 Fisher et al., 2010), often at a specific spatial, temporal, or process-level scale, depending on the
213 question of interest (Ostle et al., 2009). Some ecosystem models were later linked with TBMs in
214 order to understand vegetation patterns under current and future conditions (Pan et al., 2002).

215
216 In order to represent the interaction of roots with aboveground plant parts and the surrounding
217 soil environment (Fig. 1), ecosystem models must represent the functional balance of carbon
218 partitioning belowground to root growth, the distribution of roots throughout the soil, active root
219 functions, and the changes in partitioning and root distribution in response to changing
220 environmental conditions (Grant, 1998). Accurate model representation of root function and its
221 importance to land surface fluxes of carbon, water and nutrients is dependent on *how many* roots
222 there are, *where* roots are in the soil profile, and *which* roots are active. Unfortunately, the
223 different approaches taken with plant- and ecosystem-scale models appear to have created a gap
224 through which the representation of roots, and in particular, root function, has fallen. Some
225 ecosystem-scale process models and TBMs do not explicitly represent fine roots (Hanson et al.
226 2004), while in others, root representation is cursory, or solely to extract water from the soil.
227 Figure 4 describes model inclusion of various root processes, including root production and
228 structure, and if structure is linked to water or nutrient uptake.

229

230 In ecosystem models, plant water and nutrient uptake is usually empirically-derived from
231 functional or allometric drivers rather than mechanistically propagated based on tissue function
232 and energy expenditures (Hopmans & Bristow, 2002). N uptake from the soil profile is rarely
233 modeled in a way that depends on root properties (Table 2), although for some models N uptake
234 requires respiratory energy (Hopmans & Bristow, 2002, Fisher et al. 2010) that indicates linkages
235 to C partitioning belowground to fulfill root demand. Mycorrhizae have a large role in nutrient
236 acquisition by plants but their inclusion in root models is rare, although they are explicitly
237 represented in the detailed *ecosys* model (Grant, 1998), and implicitly represented in the *Fixation*
238 *and Uptake of Nitrogen* (FUN) root module as an extension of the root system (e.g., Fisher et al.,
239 2010), and now explicitly represented in FUN 2.0 (Brzostek et al. in review).

240
241 There are several distinct types of ecosystem models that vary in their treatment of root function:

242
243 (1) Simple modules focused on one aspect of the ecosystem that might be incorporated into
244 TBMs. For example, the Radix model estimates growth and turnover for various root classes in
245 context of internal C partitioning (Riley et al., 2009; Gaudinski et al. 2010) – such a model might
246 be leveraged to allow water and nutrient uptake dynamics from roots of different functional ages.
247 Another module, the FUN model simulates N availability and uptake based on internal C and N
248 availability, root microbial associations, water use and environmental conditions (Fisher et al.,
249 2010). This N module includes passive and active ion uptake kinetics, requiring substantial
250 respiratory energy. The model framework applies detailed ecophysiological processes to simulate
251 N uptake and internal cycling. FUN can be run as a stand-alone module or applied within TBMs
252 (e.g., JULES; Fisher et al., 2010), and ongoing work will leverage FUN into additional TBMs
253 including CLM 4.5, Noah-MP and LPJ.

254
255 (2) Whole-ecosystem models that vary in the complexity of their representation of ecosystem
256 processes (e.g., *Ecosys* (Grant 1998), G'DAY (McMurtrie et al. 2000), SPA (Williams et al.
257 1996) and TEM (Raich et al. 1991)). These four ecosystem models include representation of a
258 range of root-specific processes, based in large part on the initial ecosystem and questions

259 devised by the developers (detailed in Table 2). The models include the highly complex *ecosys*
260 model that has detailed root architecture, production and mycorrhizal colonization that can
261 respond to changing water and nutrient availability (Grant, 1998). Root water uptake in *ecosys* is
262 a function of water content, and root radial and axial resistances – the latter allows for expression
263 of dynamic root function (resistance) that can control water uptake (Grant, 1998). The *ecosys*
264 model can also differentiate nitrogen sources ($\text{NH}_4\text{-N}$ and $\text{NO}_3\text{-N}$) and includes phosphorus (P)
265 cycling, whereas most other models focus solely on nitrogen. At the opposite end of the
266 spectrum, the TEM model operates at coarse temporal and spatial scales, with focus on C and N
267 balance in soils and vegetation (Reich et al. 1991) (Table 2). There are no roots or root functions
268 present in the model. Water use is based on a water balance sub-model that includes broad site
269 characteristics including vegetation type, soils and climate. N uptake is based primarily on
270 availability, and C:N uptake costs.

271
272 (3) Optimization models attempt to avoid the pitfalls of extensive parameterization (e.g., May,
273 2004) by focusing on a few analytic expressions. One example is *MaxNup*, which optimizes the
274 vertical distribution of root biomass throughout the soil profile to maximize annual N supply to
275 aboveground plant organs (McMurtrie et al., 2012). This type of annual optimization is apparent
276 in other ‘demand’ based models, which provides a limited framework for addition of root
277 functional dynamics.

278

279 4. Terrestrial biosphere models (TBMs)

280

281 TBMs were designed to be linked into Earth system models to provide broad predictive
282 capabilities of C cycling, energy balance and climate in context of shifting natural and
283 anthropogenic forcing of the system. As with ecosystem models, TBMs must align select
284 mechanistic processes into a framework that is conducive for scaling, relying on bulk, landscape-
285 level ecosystem components and fluxes (Fig. 3). Roots, when present in a model, must be scaled
286 up from empirical data collected for specific species, or the relevant plant functional types (PFT).

287

288 Constrained by the structure of TBMs, root distribution must be represented in a single vertical
289 dimension, generally as the proportion of root mass in each of a number of soil layers, or simply
290 as a maximum rooting depth. These tend to be fixed parameters which do not exhibit dynamic
291 functionality. Root function is not usually linked with root biomass. There are some exceptions
292 such as O-CN (Zaehle & Friend, 2010) and LPJ-GUESS (Smith et al. 2013) that allow root
293 biomass to be dynamic, although even in those models, the fraction of *functional: non-functional*
294 root biomass is not dynamic. Table 3 describes how 10 commonly-used TBMs represent root
295 distribution, water and nutrient uptake.

296
297 **Water uptake in TBMs** As in the ecosystem models, water uptake in TBMs operates at the
298 macroscopic scale, determined by supply and demand. Uptake is described by a sink term in the
299 volumetric mass balance (Raats, 2007) rather than explicitly simulating the root-soil interface as
300 described in the *single root* and *individual plant* scale model sections. Plant water demand is
301 calculated as a function of atmospheric vapor pressure deficit and a series of water transport
302 resistances caused by stomata, leaf and atmospheric boundary layers, and in some cases
303 includes modeled root and stem resistances (Table 3) (e.g., SPA (Williams et al. 2001), CLM4.5
304 (GB Bonan, unpublished)). When sufficient water is available, water uptake is simulated based
305 on the plant water demand with rooting distribution or absolute rooting depth used to determine
306 the location within the soil column of water taken up by the plant. Substantial amounts of data on
307 global root distributions are available (e.g., Jackson et al. 1996, Schenk and Jackson 2002), and
308 root distribution is the most widely included root component in TBMs.

309
310 When insufficient water is available to meet demand, TBMs model uptake as a function of water
311 supply, rather than allowing for mechanistic reduction in root conductivity. Most often, supply
312 limited uptake is simulated by multiplying physiological variables with a soil water stress scalar
313 (0-1, often referred to as β), which serves to reduce demand (Feddes et al. 1978, Verhoef and
314 Egea 2014). The ' β ' soil water limitation factor can be represented as a piecewise linear function
315 of soil water matric potential, matric potential at wilting point (e.g., $\psi_{wp} = -1.5$ MPa) and matric
316 potential at a critical point below which supply limitation begins (e.g., $\psi_{fc} = -0.033$ MPa). Some

317 TBMs (e.g. CLM, Oleson et al. 2010) simulate β as a function of matric potential in relation to
318 when stomata are fully open or closed, while others (e.g. JULES (Clark et al. 2011) and CABLE
319 (Wang et al., 2010)) simulate β as function of soil water content (θ). Due to the strongly non-
320 linear relationship between ψ and θ (soil water retention curves), the two formulations allow for
321 very different supply limitation of soil water uptake. In addition, since the retention curves can
322 vary dramatically within a single profile due to changes in soil physical characteristics, relative
323 soil water availability for heterogeneous soils is not well expressed by a single relationship
324 (Warren et al. 2005), indicating a need for model parameterization of multiple soil layers
325 simultaneously where data exist.

326

327 The β term has a direct link to water uptake, thus is an obvious avenue for novel introduction of
328 dynamic root function in future TBMs. Various alternate formulations of β exist (reviewed by
329 Verhoef and Egea 2014). One of the most interesting is the inclusion of root:shoot chemical
330 (especially abscisic acid; ABA) and hydraulic signaling to control stomatal aperture and thereby
331 regulate root water uptake (Dewar 2002, Verhoef and Egea 2014). Inclusion of this ABA-based
332 water stress function provided the best fit to experimental data, although it requires additional
333 and accurate soil and plant parameter datasets – data not readily obtained at the landscape scale,
334 which limits the application and refinement of this function in TBMs. Another expression of β
335 allows for a decrease in root function under saturated, hypoxic conditions due to oxygen
336 limitation in the rhizosphere (Feddes et al. 1978), though most TBMs only consider a reduction
337 in root function in response to drying soils.

338

339 **Nitrogen uptake in TBMs** Root nitrogen (N) uptake in TBMs is also simulated at the
340 macroscopic scale by using available soil N concentrations. N uptake is simulated primarily as a
341 function of supply and often demand, as in CLM or CABLE (Thornton et al., 2007; Wang et al.,
342 2010), though the implementation varies across models far more than the implementation of
343 water uptake. Most TBMs integrate soil carbon and N cycling throughout the entire soil profile,
344 thus N uptake is from bulk soil regardless of root or N distributions within the profile, although

345 new multi-layer biogeochemical cycling algorithms are becoming available for some models
346 (e.g. CLM4.5; Koven et al., 2013).

347
348 Some TBMs use root mass as a proxy for root length density, and formulate N uptake as a linear
349 function of root mass (e.g., LM3 (Gerber et al., 2010), LPJ-GUESS (Smith et al., 2013) and O-
350 CN (Zaehle & Friend, 2010)). The linear dependence of N uptake on root mass contrasts with the
351 optimality formulation of McMurtrie et al. (2012), whereby a saturating relationship of N uptake
352 to root mass results from over-lapping nutrient depletion zones vertically within the soil profile
353 as root mass increases. Models use of biomass only, without knowledge of root anatomical or
354 functional distribution has limited ability to indicate differences between species within a plant
355 functional type (PFT). Linking biomass to function through structure is thus a key area for
356 improvement.

357
358 The LM3 and O-CN models employ a Michaelis-Menten kinetic function of N uptake, but one
359 that saturates as N supply increases. Thomas et al. (2013) modified the N dynamics of CLM4,
360 improving model accuracy at simulating N addition experiments. They showed that a key model
361 development leading to the improvement was the implementation of Michaelis-Menten kinetics
362 saturating with N supply and linearly dependent on root mass.

363
364 A number of models, e.g., LPJ-GUESS (Smith et al., 2013), O-CN (Zaehle & Friend, 2010),
365 CLM4 (Thomas et al. 2013), also simulate N uptake as a function of temperature to account for
366 the effect of temperature on metabolic rates. However, none of the models surveyed simulate N
367 uptake as a function of soil water content despite the importance of water for rhizosphere
368 nutrient cycling, for mass flow and diffusion of N to the root surface (de Willigen and van
369 Noordwijk, 1994; Cardon et al. 2013), and for oxygen dependence of metabolic rates.

370
371 **Root production in TBMs** Root growth, production and activity are dependent on carbon
372 partitioning belowground. There are a variety of different approaches to model C partitioning
373 within plants (Table 3) (Franklin et al. 2012). One promising approach (functional balance)

374 recently best represented temperate forest carbon partitioning in two Free Air CO₂ Enrichment
375 (FACE) experiments (DeKauwe et al. 2014). Functional balance approaches partition carbon to
376 various tissues to balance resource acquisition (Franklin et al., 2012), thus mechanistic model
377 improvements to allow root functional nutrient or water uptake would be dependent on
378 partitioning of carbon belowground. Representation of root function will also be necessary to
379 implement optimization schemes for partitioning in TBMs, similar to that developed by
380 McMurtrie & Dewar (2013). Flexible partitioning schemes allow vegetation turnover to vary due
381 to the different turnover times of different tissues.

382
383 Model inclusion of carbon allocation through roots to mycorrhizae and exudates may be a
384 parameter that could allow model plasticity of belowground functional dynamics, since these
385 rhizosphere processes have direct linkages to water and nutrient uptake and carbon cycling. For
386 example, observed increases in N uptake in response to elevated CO₂ were not explained by 11
387 ecosystem models (Zaehle et al. 2014) suggesting the need for additional processes by which
388 plants can stimulate N uptake through expanded effective root surface area, deeper soil mining
389 (Iversen et al. 2010, McMurtrie et al. 2012) and 'priming' of nutrient cycling (Drake et al. 2011,
390 Cheng et al. 2014). Focused root 'modules' incorporated into TBMs may allow a pathway for
391 dynamic root allocation and uptake. Indeed, the FUN nitrogen fixation module indicates
392 increased root production under elevated CO₂ FACE studies (J. Fisher, personal communication),
393 in agreement with observations, while balancing the C cost of root N uptake with other
394 respiratory and growth demands.

395
396 **Integration of detailed soil hydrologic and biogeochemical transport models into TBMs**
397 While ecosystem models and TBMs were developed with a strong plant functional component,
398 there has also been significant model development of sub-surface reactive transport dynamics in
399 the absence of vegetation (and roots). Modeling unsaturated water flow within the vadose zone is
400 achieved by mathematical approximations of one- three dimensional Richard's equations (similar
401 in structure to Darcy's Law describing saturated flow in soils and plant xylem). More recently
402 root water extraction has been added as a sink term into these detailed, highly computational

403 numerical models (Vrugt et al. 2001; Javaux et. al. 2008), which allows them to be linked into
404 TBMs. In these sub-surface hydrology models the flow of water from soil to root xylem ‘tubes’
405 is often modeled as simple one dimensional radial flow (Amenu and Kumar 2008; Schneider et.
406 al 2010), although since hydraulic conductivity changes at the soil-root interface (e.g., Carminati
407 et al. 2010) more accurate models have included an interfacial conductivity within the
408 rhizosphere (e.g., Katul et al. 2012). Modeling efforts that include rhizosphere resistance as a
409 microscopic soil-root hydraulic conductivity drop function can improve modeled dynamics of
410 water transport into roots, while actually reducing the computational time (Schroder et. al, 2008,
411 2009).

412
413 There are encouraging efforts to pair these detailed numerical reactive transport models with
414 vegetation models at the landscape level. The models have primary focus on improving surface
415 and subsurface hydrological components and often include detailed soil characteristics,
416 topography and differential water table depths (e.g., Rihani et al. 2010, Shi et al. 2013).
417 Sivandran and Bras (2013) implemented multi-layered dynamic root distribution within a
418 vegetation model (VEGGIE) coupled with a hydrologic model (tRIBS). The model dynamically
419 allocates carbon to roots at different soil layers to maximize transpiration. Simulations agreed
420 with catchment data at hourly timescales, indicating the utility for inclusion of detailed numerical
421 models in TBMs. PIHM (Qu and Duffy 2007) is a fully coupled 2-D hydrological model that has
422 been validated with extensive data at the Shale Hills Critical Zone Observatory and paired with a
423 land surface model based on the Noah LSM (Shi et al. 2013). These models include root
424 biomass-weighted water extraction by layer, and successfully simulate soil hydraulic parameters
425 and watershed discharge. Another reactive transport model, PFLOTRAN (Mills et al. 2007) has
426 been specifically designed to scale 3-D numerical hydrological modeling using parallel
427 supercomputing. PFLOTRAN is currently being linked to the CLM TBM to achieve fully-
428 coupled detailed hydrological dynamics at the land surface scale. Despite a similar lack of root
429 functional attributes in these hydrological models, they greatly improve mechanistic modeling of
430 the subsurface environment, which allows for expanded knowledge of spatial dynamics of water
431 availability. In turn, roots overlaid across the heterogeneous two-dimensional grids or three-

432 dimensional voxels in these models could be allowed step-wise increases in dynamic
433 functionality, which would greatly expand their role as a critical control point in subsurface and
434 surface ecosystem functions. The coupling of detailed subsurface models with TBMs is expected
435 continue to evolve as computational limitations diminish.

436

437 **III. Recommendations for leveraging root knowledge into models**

438

439 We have shown that there are a number of existing root models and many known root functions
440 that could be used to better represent the role of roots within TBMs. While high-resolution
441 spatial and temporal dynamics of individual roots may not be amenable for application to TBMs,
442 inclusion of specific mechanistic processes is critical to establishing a process-based
443 representation of root functionality that can be used to improve predictive capacity. Key root
444 functions that should be included in future model development include root water and nutrient
445 uptake, and carbon partitioning belowground to production, respiration, exudates and turnover.
446 Knowledge of root traits related to these functions (e.g., morphology, chemistry, mycorrhizal
447 associations) will allow those functions to be scaled into TBMs (Fig 3). Specifically, knowledge
448 of root architectural display and distribution, proportion of highly-active ephemeral or less active
449 woody roots (i.e., based on diameter, length, order, age), mycorrhizal associations, and root
450 production and turnover should be included. While some of these parameters are already
451 included in TBMs, most are not well represented (e.g., Fig. 4), indicating dynamic functionality
452 could be improved or added. Dynamics to consider include plasticity of roots to environmental
453 conditions - especially increased root water and nutrient uptake kinetics and root proliferation in
454 resource rich areas, and reduction in root activity in resource poor areas. These dynamics should
455 be linked to spatial and temporal changes in environmental conditions through both theoretical
456 and empirical studies that intersect process- and trait-based parameterization.

457 Unfortunately, there is not a good understanding of TBM model sensitivity to root
458 function; i.e., if inclusion of mechanistic root functions in models could improve model
459 performance within the current model framework, although studies that have included more root

460 parameters have yielded better results (e.g., inclusion of dynamic root area (Schymanski et al.,
461 2008) or hydraulic redistribution (Lee et al. 2005)).

462
463 In the following section we assess how our current mechanistic knowledge of root function
464 interacts with and determines ecosystem function, and suggest what should be taken into
465 consideration when modeling roots in TBMs. Areas of discussion include root distribution and its
466 utility for scaling, linking root traits to root functions, key regulatory factors such as water
467 uptake kinetics (including hydraulic redistribution) and nutrient uptake kinetics, data availability,
468 and strategies for model improvement. Figure 5 provides a framework for root data and model
469 assessment, and how we might proceed towards improved models or novel stand-alone root
470 modules that could be embedded within TBMs.

471

472 1. Scaling root function using root architecture

473

474 Root distribution within the soil profile provides the basic foundation for root function, and is the
475 characteristic most frequently included in large-scale TBMs as a regulator of water uptake (Fig 4,
476 5). Data are widespread and readily obtained destructively through soil coring and excavation
477 (e.g., Nadezhdina and Cermak 2003), or through in situ observations (rhizotrons, minirhizotrons)
478 (Pierret et al. 2005, Iversen et al. 2012). Specific root structural traits can then be overlaid on this
479 distribution, with allowance for environmental gradients and biotic signals to shift trait functions
480 within that distribution (Fig 5). For example, during a period when upper soils dry, the upper
481 roots become less functional, only to rapidly increase in function following precipitation inputs
482 (e.g. Warren et al. 2005). Root proliferation can decrease total root system hydraulic resistance
483 under environmental stress, increasing capacity for water uptake and increasing the root:shoot
484 ratio (Steudle 2001). Inclusion of a dynamic root:shoot ratio in TBMs could bound C and water
485 flux at the landscape level for a specific set of resources, as demonstrated with a plant scale
486 model by Sperry et al. (1998).

487

488 Shifts in actual or *functional* root distributions within the soil profile represent a dynamic
489 functionality of the root system that is difficult to include in TBMs, although several research
490 directions linked to root function are quite promising, including linking function to root class and
491 characteristic root traits, and consideration of water stress and hydraulic redistribution through
492 the soil profile (e.g., Valenzuela-Estrada et al. 2008). For example, Schymanski et al. (2008) used
493 an optimality function to meet canopy demands for water uptake by allowing root surface area to
494 be dynamic and thereby able to shift into moister soil as necessary. The model ran on a one day
495 time step, and while this may not accurately represent new root growth, it does represent shifts in
496 root functionality within an existing root system. Results including this dynamic functionality
497 improved estimates of water flux from a tropical savanna as compared with a static root system.
498 Inclusion of such plasticity of root function provides a significant step toward better mechanistic
499 representation of roots in models that could improve model performance.

500

501 Different plant functional types (PFTs) vary in root display (presence of taproot, lateral spread,
502 dimorphism), maximum depth, and morphological traits that affect their interaction with the soil
503 (Canadell et al. 1996, Schenk 2005, Pohl et al. 2011). Root distribution varies across biomes and
504 does not necessarily depend on soil depth. A global synthesis indicates mean maximum rooting
505 depths range from 2.6 m for herbs to 7.0 m for trees (Canadell 1996); although root distributions
506 across biomes tend to be only as deep as necessary to supply evapotranspirational demand,
507 allowing prediction of community root distribution based primarily on precipitation and potential
508 evapotranspiration (Schenk 2008). While simplified distributions of roots are readily
509 incorporated into models, Feddes (2001) suggested the need to continue modeling efforts from a
510 bottom up mechanistic approach, as well as a top-down approach, in order to provide process-
511 level understanding to these simplified models.

512

513 2. Linking root function to traits

514

515 Plant species responses to resource availability vary due to differences in competitive strategies
516 (Hodge et al. 2004). In context of drought, some species have adapted growth of deep roots to

517 tap groundwater (Meinzer 1927), in some cases up to 50 m (Canadell 1996), while others with
518 shallower root systems close stomata to limit water use and tolerate arid conditions. Such
519 variation reiterates the necessity to include root traits within plant functional type (PFT)
520 classifications in order to adequately scale functionality of root architecture into the models. At
521 the landscape scale, the distribution of root traits, specialized root structures (cluster roots, root
522 hairs) and mycorrhizal associations reflect resource availability (Lambers et al. 2008). Root
523 function can be linked to characteristic root traits that vary across species (e.g., Comas and
524 Eissenstat 2009, Kong et al. 2014) and PFTs (especially annual versus perennial), although other
525 than root distribution, few, if any root traits are included in PFT classifications (Wullschleger et
526 al. 2014), or TBMs. Currently, TBMs use static plant parameters for each PFT, even though
527 phenotypic expression of traits is strongly affected by variations in environmental conditions;
528 inclusion of photosynthetic traits that were allowed to vary linearly with climate within PFTs
529 shifted simulated biomass estimates and PFT cover-type by 10-20% for forests compared with
530 the default simulations (Verheijen et al. 2013). Root turnover rates are a key root trait linked to
531 ecosystem function that can have substantial variation across species within PFT; modeled inter-
532 species shifts in root turnover within PFT under climate change had substantial implications at
533 the landscape level (McCormack et al. 2013). Efforts to understanding gene linkages to turnover
534 and other root traits provide a pathway for screening of individual species' root characteristics,
535 an effort particularly advanced for crop systems where traits are being linked to gross primary
536 production and drought resistance (Comas et al. 2013). Further phenotyping research is required
537 in natural ecosystems to create the database necessary for inclusion of variable, dynamic root
538 traits into TBMs. A trait-based, mechanistic representation of roots in TBMs will have significant
539 impacts on model outputs.

540

541 Key root functional traits to consider for models are root morphology, chemistry and microbial
542 associations, since they control dynamics of water and nutrient ion flux through the soil into
543 roots under varying environmental conditions (Figs 1, 5). The white, ephemeral first and second
544 order roots are the predominant pathway for water and nutrient uptake (Steudle 2000, Guo et al.
545 2008, Rewald et al. 2011), although coarser suberized woody roots also provide a persistent, yet

546 lower uptake pathway that may be important for seedlings (Hawkins et al. 2014), or seasonally
547 during periods of low fine root growth or activity (Van Rees and Comerford 1990, Lindenmair et
548 al. 2004), and which may be associated with sustained root rhizosphere hydration through
549 hydraulic redistribution (Rewald et al. 2011). Root hairs and mycorrhizal associations can
550 enhance the effective surface area of the root system and increase the potential for resource
551 extraction in many species (Read & Boyd 1986; Augé, 2001, Segal et al. 2008).

552
553 Refinement of the ‘fine:coarse’ root ratios used in some models should reflect root function, not
554 just root size, which varies by species. Root orders, and their function can be characterized
555 indirectly by relative degree of mycorrhizal colonization, root density or root C:N ratio
556 (Valenzuela-Estrada et al. 2008). Root lifespan is another key root attribute that might be
557 correlated with these and other root traits such as diameter, depth (Pritchard and Strand 2008),
558 specific root length (McCormack et al. 2012) or root and aboveground traits together (root
559 diameter and plant growth) as found in twelve temperate tree species (McCormack et al. 2012).
560 Knowledge of root traits can be used to improve models of water or nutrient uptake kinetics
561 (e.g., refining active root absorbing area, or classifying root function in the FUN N uptake
562 module), add functionality to existing modules of root turnover (e.g., Radix), and to provide
563 scalable trait data for novel root functional representation in TBMs (Fig. 3).

564
565 3. Water uptake

566
567 The process of root water uptake includes some regulatory steps that could be included in TBMs.
568 Under moist soil conditions, radial resistance limits root water uptake and is actively controlled
569 by membrane bound transport proteins (aquaporins) that respond to osmotic gradients
570 (Chrispeels et al. 1999, Steudle 2000, Aroca et al. 2012). Under drying conditions water uptake is
571 regulated by varying soil and plant resistances to water movement (Blizzard and Boyer 1980,
572 Sperry et al. 1998, Hacke et al. 2000). Radial hydraulic conductivity through aquaporin
573 regulation can be rapidly increased or decreased based on perceived environmental stimuli
574 including mycorrhizal colonization (Lehto and Zwiazek 2011) or suboptimal environmental

575 conditions (e.g., drought, temperature, anoxia; Siemens and Zwiazek 2004). Indeed, deep roots
576 in wet soils upregulated aquaporins during drought, increasing hydraulic conductivity
577 substantially as shallow root conductivity declined (Johnson et al. 2014). Root stress responses
578 are often reflected in production and accumulation of abscisic acid (ABA) or other plant growth
579 regulators (Davies and Zhang 1991; Wilkinson and Davies 2002; Aroca et al. 2012). Root
580 derived plant regulators or mycorrhizal-derived inorganic ions can be transported through the
581 xylem to elicit a response in the leaves, particularly stomatal closure (Davies et al. 1994).
582 Similarly, two-way hydraulic signaling also connects root and shoot functions allowing
583 coordinated whole plant response to changing soil or atmospheric conditions (e.g., Blackman and
584 Davies 1985, Comstock 2002, Meinzer 2002, Vandeleur et al. 2014). Pathway resistances are
585 included in some TBMs, however, none to our knowledge have active regulation based on
586 aquaporin expression, which could provide a mechanistic control on water use and improve
587 model performance, similar to application of a dynamic ABA parameter on the water stress
588 scalar, β , as described earlier. β is an obvious target for providing dynamic, albeit indirect,
589 functionality to water uptake since it already exists in many models, and would be particularly
590 useful if weighted by root *functional* class (e.g., age, order, morphology) within each soil layer.

591

592 4. Hydraulic redistribution

593

594 Hydraulic redistribution (HR) can maintain fine root function (Domec et al. 2004), extend root
595 life (Bauerle et al. 2008), rehydrate the rhizosphere (Emerman and Dawson 1996), enhance
596 nutrient availability (Cardon et al. 2013) and acquisition (Matimati et al. 2014), and should
597 prolong soil-root contact under dry conditions. HR's contribution to total site water use is known
598 to vary widely depending on the ecosystem (Neumann and Cardon 2012); yet even minor HR
599 can provide significant benefits for continued root and mycorrhizal function during drying
600 conditions. HR has been represented by variation in water transport between soil layers, dynamic
601 soil-plant-atmosphere resistances, radial/axial conductivity *big root* models, and root optimality
602 models (Neumann and Cardon 2012). Results indicate that the inclusion of HR can help explain
603 patterns of soil and plant water flux for individual trees (e.g., David et al. 2013), resulting in

604 significant implications for stand- (Domec et al. 2010) and landscape-scale (Lee et al. 2005,
605 Wang et al. 2011) carbon uptake and water release. Application to the large-scale models
606 included HR as an additional water flux term in the NCAR Community Atmospheric Model
607 Version 2 (CAM2) coupled with the Community Land Model (CLM) (Lee et al. 2005) and in
608 CLM3 coupled with a dynamic global vegetation model (CLM3-DGVM) (Wang et al. 2011).
609 Results suggest inclusion of HR can increase dry season water use in the Amazon forests by 40%
610 (Lee et al. 2005), but may exacerbate plant water stress under extended drought if soil water is
611 exhausted (Wang et al. 2011) – both efforts illustrate how a small change in root function can
612 result in substantial implication for global scale. HR is a process that should be included in large-
613 scale models, but it will require consideration of depth specific soil-plant water dynamics,
614 internal competition for water within the plant vascular system (Sperry et al. 1998), plant water
615 capacitance (Scholtz et al. 2007) and nocturnal transpiration (Caird et al. 2007, Dawson et al.
616 2007, Fisher et al. 2007, Zeppel et al. 2012) to account for concurrent uptake and release
617 dynamics (Neumann and Cardon 2012).

618

619 5. Ion uptake kinetics

620

621 Mineral ions are transported into the root cortex via mass flow, diffusion, or through mycorrhizal
622 absorption, which is particularly important for uptake of immobile nutrients such as phosphorus.
623 Movement through the plasma membrane of root endodermal cells is facilitated by a variety of
624 passive or active transport proteins, including ATP-fueled ion pumps (Chrispeels et al. 1999). Ion
625 absorption kinetics vary by species depending upon the nutrient concentration, with multiple low
626 and high affinity mechanisms controlled by environmental conditions (Epstein 1966, Chapin
627 1980, Chrispeels et al. 1999, BassiriRad 2000). Root nutrient uptake kinetics are often measured
628 on intact or excised roots under well hydrated conditions, i.e., not under water stress. In drought
629 tolerant woody sagebrush, nitrogen and phosphorus uptake rates were maintained or even
630 increased under laboratory water potential stress, illustrating the uncoupling of water and
631 nutrient flux into the root (Matzner and Richards 1996). Under drying conditions, *in situ* nutrient
632 absorption does not appear to be limited by uptake kinetics, but rather by diffusion of ions

633 through the soil to the root surface (Chapin 1980). Mycorrhizae can span soil-root gaps and help
634 to maintain a viable transport pathway from soil to root under drying conditions.

635

636 Absolute uptake kinetics for specific ions are thus a function of a variety of control points.
637 Improved mechanistic representation of ion uptake in models will require inclusion and
638 expanded consideration of Michaelis–Menten kinetics used in some TBMs (Fig. 4). One key
639 improvement would be to allow the kinetics to vary by depth in response to environmental
640 conditions such as temperature or soil water content (i.e., through the β stress scalar), weighted
641 by specific root traits and root functional classes. Root hydraulic conductivity (i.e., aquaporin
642 function) is often upregulated by soil ion concentrations such as nitrate, resulting in whole plant
643 hydraulic signaling (Gorska et al. 2008, Cramer et al. 2009), increased root uptake kinetics
644 (Jackson et al. 1990) and proliferation of roots in resource rich areas (reviewed in Hodge et al.
645 2004). Such plasticity in function might require a multicomponent ion uptake kinetic model that
646 includes the appropriate regulatory and substrate parameters. One modeling framework to
647 consider involves a modification of the HYDRUS reactive transport model. The model was
648 modified to allow a ‘root adaptability factor’ which compensates for reduced water and nutrient
649 uptake by stressed roots in resource poor areas by increasing uptake of roots in unstressed soil
650 (Šimůnek and Hopmans 2009). Such efforts to refine existing models through use of dynamic
651 scalars allows improved approximation of the processes inherent in more complex models,
652 without the necessity of novel modeling frameworks and collection of additional data.

653

654 6. Available Root data – a Serious Limitation

655

656 A fine balance exists between accurately representing ecological processes, and the added
657 uncertainty that comes with model complexity in terms of appropriate and accurate
658 parameterization, which may require regional or global data sets (Fisher et al., 2010). A
659 concentrated effort needs to be made to fill the gaps in the trait database to obtain accurate
660 representation of the trait space of terrestrial plants and ecosystems. There is a need for
661 development of databases across PFTs of both root distribution, root structure and root functional

662 traits that are linked to specific plant responses to environmental conditions. Recent investigation
663 of root traits of 96 subtropical angiosperm trees illustrates the broad variation and plasticity in
664 traits within a single PFT (Kong et al. 2014), as well as the necessity to identify trait covariance
665 and linkages to function (Iversen 2014). Key root traits to compile into databases include length,
666 diameter, order, display, age, C:N and mycorrhizal associations.

667
668 A wealth of belowground datasets exist globally – including detailed soil and physical
669 characteristics (described in Feddes 2001), and estimates of minimum, mean and maximum
670 rooting depths (e.g., Canadell 1996; Schenk and Jackson 2002) and root biomass, length and
671 nutrient content (Jackson et al. 1997) for different biomes. Characteristics of the root system
672 most amenable to use in TBM's include root biomass, depth distribution, production and
673 turnover, fine:coarse root ratios and nutrient content (Feddes 2001). Information on dynamic root
674 functioning under varied environmental conditions, however, remains disparate, non-
675 standardized and dispersed. Certainly, there is an immense amount of data regarding root
676 phenotypic plasticity to water, nutrient and temperature treatments for different species, different
677 root anatomies and at various ontogenetic stages. For future application to TBMs, root functional
678 data should be linked with scalable root traits whenever possible (Iversen 2014), including
679 covariate plant traits (e.g., height, leaf area)(McCormack et al. 2012, Wullschleger et al. 2014),
680 and correlated to concurrent data collection of environmental conditions that regulate root
681 function (e.g., root depth, soil temperature, texture, water content and nutrient availability,
682 atmospheric vapor pressure deficit, etc.)

683
684 Scaling root traits to the landscape level can be facilitated by leveraging the expansive research
685 and data derived from existing (e.g., Fluxnet, LTER, Critical Zone Observatories) and new (e.g.,
686 NEON, AnaEE) long term ecological research sites (described by Peters et al. 2014).

687 Observational studies can be nested in plots within an ecosystem (Bradford et al. 2010), within a
688 watershed (Anderson et al. 2010), or within the footprint of eddy covariance towers (Law et al.
689 2006) to provide scaling across the landscape. Such nested studies provide a valuable framework

690 to allow scaling of discrete mechanistic knowledge of root function to realized fluxes at the land
691 surface.

692

693 7. Novel modeling platforms

694

695 Many TBMs have quite complex interlinked source files and algorithms that when paired with
696 earth system models makes testing of specific mechanistic process simulations slow and difficult
697 (Wang et al. 2014). In addition, the structure is not easy to assess or comprehend by non-
698 modelers, thereby excluding experimentalists from model development and improvement efforts.
699 However, new initiatives to pull out specific functional parameters from TBMs are promising.
700 For example, a new functional testing platform has been developed for CLM (the land
701 component of the Community Earth System Model), which has successfully extracted the
702 photosynthetic sub-unit from CLM for testing and modification, and includes a user-friendly
703 GUI (Wang et al. 2014). Both extraction of belowground functional modules in current TBMs,
704 and addition of new modules (e.g. FUN, RADIX) provide a pathway for inclusion of novel or
705 refined root components that can lead to model improvements. In addition, TBMs can be run at
706 the 'point' scale, using site-specific parameters to inform model PFTs, to understand processes
707 operating in a plot or experimental manipulation (e.g., Ostle et al., 2009, De Kauwe et al., 2013;
708 Zaehle et al., 2014; Walker et al., in press).

709

710 An essential component to improve model representation of root functional processes is to
711 partition function throughout the soil profile, similar to how some models treat the leaf canopy.
712 Some TBMs are being improved to include more than energy or water dynamics in each soil
713 layer by addition of C and N dynamics through the soil profile (e.g., CLM4.5; Koven et al.,
714 2013). Root dynamics should be progressively integrated into those multilayered soil
715 formulations by moving beyond just a parameterized value of root distribution.

716

717 Specific model improvements might include the addition of spatial and temporal dynamics of
718 root production and turnover, and water/nutrient uptake kinetics linked to refined functional

719 classes of roots (i.e., based on traits such as length, diameter, order, display, age, C:N and
720 mycorrhizal associations) that vary in their functional response to environmental conditions or
721 internal signals. The distribution of roots might be seasonally and annually dynamic to
722 proliferate (or upregulate function) into resource rich areas, and diminish in stressful, resource
723 poor areas (e.g., Schymanski et al. 2008). The differential root activity and turnover reflected by
724 such a model could further be linked to rhizosphere microbial carbon and nutrient cycling
725 processes.

726

727 **IV. Conclusions**

728

729 Interactions between plant roots and the surrounding soil environment (especially gradients,
730 distributions, and functions with depth) are required to accurately represent root uptake of
731 nutrients and water under changing environmental conditions, as well as plant C release to soils
732 (Grant, 1998). Current model distribution of roots is usually static and discrete and thus is not
733 representative of actual dynamic root exploration, function or turnover, nor linked to mechanistic
734 biotic and biogeochemical cycling within the rhizosphere. Despite substantial mechanistic
735 knowledge of root function, data assimilation, oversimplification and scaling issues continue to
736 limit detailed representation of roots in TBMs. Development of well-documented, error-checked
737 databases of root, soil and environmental dynamics are a priority that will be critical to porting
738 mechanistic function into TBMs – key examples include the successful plant trait-based *TRY*
739 (Kattage et al. 2011) and photosynthetic *LeafWeb* (Gu et al. 2010) databases. Emphasis should be
740 placed on assessing model sensitivity to root processes, then development and refining the root
741 modules and functional testing platforms that can lead to improved mechanistic representing of
742 root processes in TBMs (Fig. 5). Promising root processes that might be included in future
743 modeling activities include addition of dynamic root distribution, production and turnover,
744 proportions of highly active, ephemeral roots, mycorrhizal associations, dynamic water and ion
745 extraction, and hydraulic redistribution. Paired with new data compilation efforts, new model
746 tools, and new model development, the representation of roots in TBMs is expected to continue

747 to evolve and lead to advances in predictive capacity of carbon, water and energy fluxes at the
748 land surface.

749

750 **Acknowledgements**

751

752 The authors appreciate comments from Richard Norby, Josh Fisher, and two anonymous
753 reviewers, as well as editorial assistance by Terry Pfeiffer. This material is based upon work
754 supported by the U.S. Department of Energy, Office of Science, Office of Biological and
755 Environmental Research, under contract DE-AC05-00OR22725.

756

757 **References**

758

759 **Agren GI, McMurtrie RE, Parton WJ, Pastor J, Shugart HH. 1991.** State-of-the-art of
760 models of production decomposition linkages in conifer and grassland ecosystems.

761 *Ecological Applications* **1**: 118-138.

762 **Allen MF, Kitajima K. 2013.** In situ high-frequency observations of mycorrhizas. *New*
763 *Phytologist* **200**: 222-228.

764 **Amenu GG, Kumar P. 2008.** A model for hydraulic redistribution incorporating coupled soil-
765 root moisture transport. *Hydrology and Earth System Sciences* **12**: 55–74.

766 **Anderson RS, Anderson S, Aufdenkampe AK, Bales R, Brantley S, Chorover J, Duffy CJ,**
767 **Scatena FN, Sparks DL, Troch PA, Yoo K. 2010.** Future directions for critical zone
768 observatory (CZO) science. CZO Community, December 29, 2010.

769 **Aroca, R, Porcel R, Ruiz-Lozano JM. 2012.** Regulation of root water uptake under abiotic
770 stress conditions. *Journal of Experimental Botany* **63**: 43-57.

771 **Augé RM. 2001.** Water relations, drought and vesicular-arbuscular mycorrhizal symbiosis.
772 *Mycorrhiza* **11**: 3-42.

773 **Band LR, Fozard JA, Godin C, Jensen OE, Pridmore T, Bennett MJ, King JR. 2012.**
774 Multiscale systems analysis of root growth and development: modeling beyond the
775 network and cellular scales. *The Plant Cell* **24**: 3892–3906.

- 776 **Barber SA. 1962.** A diffusion and mass-flow concept of soil nutrient availability. *Soil Science*
777 **93:** 39-49.
- 778 **BassiriRad, H. 2000.** Kinetics of nutrient uptake by roots: responses to global change. *New*
779 *Phytologist* **147:** 155-169.
- 780 **Bauerle TL, Richards JH, Smart DR, Eissenstat DM 2008.** Importance of internal hydraulic
781 redistribution for prolonging lifespan of roots in dry soil. *Plant Cell and Environment* **31:**
782 171-186.
- 783 **Bingham IJ, Glass ADM, Kronzucker HJ, Robinson D, Scrimgeour CM. 2000.** Isotope
784 Techniques. In “Root Methods”. Edited by Smit AL et al. Springer Berlin Heidelberg, pp
785 365-402
- 786 **Blackman PG, Davies WJ. 1985.** Root to shoot communication in maize plants of the effects of
787 soil drying. *Journal of Experimental Botany* **36:** 39-48.
- 788 **Blizzard WE, Boyer JS. 1980.** Comparative resistance of the soil and the plant to water
789 transport. *Plant Physiology* **66:** 809–814.
- 790 **Bonan GB, Lawrence PJ, Oleson KW, Levis S, Jung M, Reichstein M, Lawrence DM,**
791 **Swenson SC. 2011.** Improving canopy processes in the Community Land Model version 4
792 (CLM4) using global flux fields empirically inferred from FLUXNET data. *Journal of*
793 *Geophysical Research-Biogeosciences* **116:** G02014.
- 794 **Bradford JB, Weishampel P, Smith M-L, Kolka R, Birdsey RA, Ollinger SV, Ryan MG.**
795 **2010.** Carbon pools and fluxes in small temperate forest landscapes: Variability and
796 implications for sampling design. *Forest Ecology and Management* **259:** 1245-1254.
- 797 **Brzostek ER, Fisher JB, Phillips RP. 2014.** Modeling the carbon cost of plant nitrogen
798 acquisition: mycorrhizal trade-offs and multi-path resistance uptake improve predictions of
799 retranslocation. *Journal of Geophysical Research – Biogeosciences (under review)*
- 800 **Busch W, Moore BT, Martsberger B, Mace DL, Twigg RW, Jung J, Pruteanu-Malinici I,**
801 **Kennedy SJ, Fricke GK, Clark RL et al. 2012.** A microfluidic device and computational
802 platform for high-throughput live imaging of gene expression. *Nature Methods* **9:** 1101-
803 1106.

- 804 **Caird MA, Richards JH, Donovan LA. 2007.** Nighttime stomatal conductance and
 805 transpiration in C-3 and C-4 plants. *Plant Physiology* **143**: 4-10.
- 806 **Canadell J, Jackson RB, Ehleringer JR, Mooney HA, Saia OE, Schulze ED. 1996.**
 807 Maximum rooting depth of vegetation types at the global scale *Oecologia* **108**: 583-595.
- 808 **Cardon ZG, Stark JM, Herron PM, Rasmussen JA. 2013.** Sagebrush carrying out hydraulic
 809 lift enhances surface soil nitrogen cycling and nitrogen uptake into inflorescences.
 810 *Proceedings of the National Academy of Sciences, USA* (in press)
 811 *doi:10.1073/pnas.1311314110*.
- 812 **Carminati A, Moradi AB, Vetterlein D, Vontobel P, Lehmann E, Weller U, Vogel H, Oswald**
 813 **SE. 2010.** Dynamics of soil water content in the rhizosphere. *Plant and Soil* **332**: 163-176
- 814 **Chapin FS III. 1980.** The mineral-nutrition of wild plants. *Annual Review of Ecology and*
 815 *Systematics* **11**: 233-260.
- 816 **Cheng W, Parton WJ, Gonzalez-Meler MA, Phillips R, Asao S, McNickle GG, Brzostek E,**
 817 **Jastrow JD. 2014.** Synthesis and modeling perspectives of rhizosphere priming. *New*
 818 *Phytologist* **201**: 31-44.
- 819 **Chrispeels MJ, Crawford NM, Schroeder JI. 1999.** Proteins for transport of water and mineral
 820 nutrients across the membranes of plant cells. *The Plant Cell* **11**: 661-675.
- 821 **Clark DB et al. 2011.** The Joint UK Land Environment Simulator (JULES), model description –
 822 Part 2: Carbon fluxes and vegetation dynamics. *Geoscientific Model Development* **4**: 701–
 823 722, *doi:10.5194/gmd-4-701-2011*
- 824 **Clausnitzer V, Hopmans JW. 1994.** Simultaneous modeling of transient three-dimensional root
 825 growth and soil water flow. *Plant and Soil* **164**: 299-314.
- 826 **Comas LH, Eissenstat DM. 2009.** Patterns in root trait variation among 25 co-existing North
 827 American forest species. *New Phytologist* **182**: 919-928.
- 828 **Comas LH, Becker SR, Cruz VMV, Byrne PF, Dierig DA. 2013.** Root traits contributing to
 829 plant productivity under drought. *Frontiers in Plant Science* **4**:442. *doi:*
 830 *10.3389/fpls.2013.00442*.
- 831 **Comstock, JP. 2002.** Hydraulic and chemical signaling in the control of stomatal conductance
 832 and transpiration. *Journal of Experimental Botany* **53**: 195-200.

- 833 **Cramer, MD, Hawkins H-J, Verboom GA. 2009.** The importance of nutritional regulation of
834 plant water flux. *Oecologia* **161**: 15-24.
- 835 **David TS, Pinto CA, Nadezhdina N, Kurz-Besson C, Henriques MO, Quilhó T, Cermak J,**
836 **Chaves MM, Pereira JS et al. 2013.** Root functioning, tree water use and hydraulic
837 redistribution in *Quercus suber* trees: A modeling approach based on root sap flow. *Forest*
838 *Ecology and Management* **307**: 136-146.
- 839 **Davies WJ and Zhang J. 1991.** Root signals and the regulation of growth and development of
840 plants in drying soil. *Annual Review of Plant Physiology and Plant Molecular Biology* **42**:
841 55-76.
- 842 **Davies WJ, Tardieu F, Trejo CL. 1994.** How do chemical signals work in plants that grow in
843 drying soil? *Plant Physiology* **104**: 309–314.
- 844 **Dawson TE, Burgess SSO, Tu KP, et al. 2007.** Nighttime transpiration in woody plants from
845 contrasting ecosystems. *Tree Physiology* **27(4)**: 561–575.
- 846 **De Kauwe MG, Medlyn BE, Zaehle S, Walker AP, Dietze MC, Hickler T, Jain AK, Luo Y,**
847 **Parton WJ, Prentice IC, et al. 2013.** Forest water use and water use efficiency at elevated
848 CO₂: a model-data intercomparison at two contrasting temperate forest FACE sites. *Global*
849 *Change Biology* **19**: 1759–1779.
- 850 **Dewar RC. 2002.** The Ball–Berry–Leuning and Tardieu–Davies stomatal models: synthesis and
851 extension within a spatially aggregated picture of guard cell function. *Plant Cell and*
852 *Environment* **25**: 1383–1398.
- 853 **de Willigen P, van Dam JC, Javaux M, Heinen M. 2012.** Root water uptake as simulated by
854 three soil water flow models. *Vadose Zone Journal* **11**: doi:10.2136/vzj2012.0018.
- 855 **de Willigen P, van Noordwijk M. 1994.** Mass flow and diffusion of nutrients to a root with
856 constant or zero-sink uptake I. Constant uptake. *Soil Science* **157**: 162-170.
- 857 **Diggle AJ. 1988.** ROOTMAP - a model of three-dimensional coordinates of the growth and
858 structure of fibrous root systems. *Plant and Soil* **105**: 169-178.
- 859 **Domec JC, King JS, Noormets A, Treasure EA, Gavazzi MJ, Sun G, McNulty SG. 2010.**
860 Hydraulic redistribution of soil water by roots affects whole stand evapotranspiration and
861 net ecosystem carbon exchange. *New Phytologist* **187**: 171-183.

- 862 **Domec J-C, Warren JM, Meinzer FC, Brooks JR, Coulombe R. 2004.** Native root xylem
 863 embolism and stomatal closure in stands of Douglas-fir and ponderosa pine: mitigation by
 864 hydraulic redistribution. *Oecologia* **141**:7-16.
- 865 **Doussan C, Pierret A, Garrigues E, Pagès L. 2006.** Water uptake by plant roots: II - Modeling
 866 of water transfer in the soil root-system with explicit account of flow within the root
 867 system - Comparison with experiments. *Plant and Soil* **283**: 99-117.
- 868 **Drake JE, Gallet-Budynek A, Hofmockel KS, Bernhardt ES, Billings SA, Jackson RB,**
 869 **Johnsen KS, Lichter J, McCarthy HR, McCormack ML, et al. 2011.** Increases in the
 870 flux of carbon belowground stimulate nitrogen uptake and sustain the long-term
 871 enhancement of forest productivity under elevated CO₂. *Ecology Letters* **14**: 349–357.
- 872 **Draye X, Kim Y, Lobet G, Javaux M. 2010.** Model-assisted integration of physiological and
 873 environmental constraints affecting the dynamic and spatial patterns of root water uptake
 874 from soils. *Journal of Experimental Botany* **61**: 2145-2155.
- 875 **Dunbabin VM, Diggle AJ, Rengel Z, van Hugten R. 2002.** Modeling the interactions between
 876 water and nutrient uptake and root growth. *Plant and Soil* **239**: 19-38.
- 877 **Dunbabin V, Diggle A, Rengle Z. 2003.** Is there an optimal root architecture for nitrate capture
 878 in leaching environments? *Plant, Cell and Environment* **26**: 835-844.
- 879 **Dunbabin V, Rengel Z, Diggle AJ. 2004.** Simulating form and function of root systems:
 880 efficiency of nitrate uptake is dependent on root system architecture and the spatial and
 881 temporal variability of nitrate supply. *Functional Ecology* **18**: 204-211.
- 882 **Dupuy LX, Fourcaud T, Lac P, Stokes A. 2007.** A generic 3D finite element model of tree
 883 anchorage integrating soil mechanics and real root system architecture. *American Journal*
 884 *of Botany* **94**: 1506-1514.
- 885 **Dupuy L, Gregory PJ, Bengough AG. 2009.** Root growth models: towards a new generation of
 886 continuous approaches. *Journal of Experimental Botany* **61**: 2131–2143.
- 887 **Emerman SH, Dawson TE. 1996.** Hydraulic lift and its influence on the water content of the
 888 rhizosphere: an example from sugar maple, *Acer saccharum*. *Oecologia* **108**: 273–278.
- 889 **Epstein E. 1966.** Dual pattern of ion absorption by plant cells and by plants. *Nature* **212**: 1324-
 890 1327.

- 891 **Euskirchen ES, McGuire AD, Chapin FS, Yi S, Thompson CC. 2009.** Changes in vegetation
892 in northern Alaska under scenarios of climate change, 2003-2100: Implications for climate
893 feedbacks. *Ecological Applications* **19**: 1022-1043.
- 894 **Feddes RA, Hoff H, Bruen M, Dawson T, de Rosnay P, Dirmeyer P, Jackson RB, Kabat P,**
895 **Kleidon A, Lilly A et al. 2001.** Modeling root water uptake in hydrological and climate
896 models. *Bulletin of the American Meteorological Society* **82**: 2797-2809.
- 897 **Feddes RA, Kowalik PJ, Zaradny H. 1978.** *Simulation of field water use and crop yield.*
898 Oxford, UK: John Wiley and Sons.
- 899 **Fisher, J.B., Baldocchi, D.D., Misson, L., Dawson, T., Goldstein, A.H. 2007.** What the towers
900 don't see at night: Nocturnal sap flow in trees and shrubs at two AmeriFlux sites in
901 California. *Tree Physiology* **27**: 597-610.
- 902 **Fisher JB, Sitch S, Malhi Y, Fisher RA, Huntingford C, Tan SY. 2010.** Carbon cost of plant N
903 acquisition: A mechanistic, globally applicable model of plant N uptake, retranslocation,
904 and fixation. *Global Biogeochemical Cycles* **24**: GB1014.
- 905 **Franklin O, Johansson J, Dewar R, Dieckmann U, McMurtrie RE, Brännström A,**
906 **Dybzinski. 2012.** Modeling carbon allocation in trees: a search for principles. *Tree*
907 *Physiology* **32**: 648-666.
- 908 **Gardner WR. 1960.** Dynamic aspects of water availability to plants. *Soil Science* **89**: 63-73.
- 909 **Gardner WR 1964.** Relation of root distribution to water uptake and availability. *Agronomy*
910 *Journal* **56**: 41-45.
- 911 **Gardner WR. 1965.** Dynamic aspects of soil water availability to plants. *Annual Review of*
912 *Plant Physiology* **16**: 323-342.
- 913 **Gaudinski, JB, Torn, MS, Riley, WJ, Dawson, TE, Joslin JD, Majdi H. 2010.** Measuring and
914 modeling the spectrum of fine-root turnover times in three forests using isotopes,
915 minirhizotrons, and the Radix model. *Global Biogeochemical Cycles* **24**: GB3029,
916 doi:10.1029/2009GB003649.
- 917 **Ge Z, Rubio G, Lynch JP. 2000.** The importance of root gravitropism for inter-root competition
918 and phosphorus acquisition efficiency: results from a geometric simulation model. *Plant*
919 *and Soil* **218**: 159-171.

- 920 **Gerber S, Hedin LO, Oppenheimer M, Pacala SW, Shevliakova E. 2010.** N cycling and
 921 feedbacks in a global dynamic land model. *Global Biogeochemical Cycles* **24**: GB1001.
- 922 **Gorska A, Ye Q, Holbrook NM, Zwieniecki MA. 2008.** Nitrate control of root hydraulic
 923 properties in plants: translating local information to whole plant response. *Plant*
 924 *Physiology* **148**: 1159-1167.
- 925 **Grant RF. 1998.** Simulation in ecosys of root growth response to contrasting soil water and N.
 926 *Ecological Modelling* **107**: 237-264.
- 927 **Gu L, Pallardy, SG, Tu K, Law BE, Wullschleger SD. 2010.** Reliable estimation of
 928 biochemical parameters from C3 leaf photosynthesis–intercellular carbon dioxide response
 929 curves. *Plant, Cell and Environment* **33**: 1852-1874.
- 930 **Guo DL, Xia MX, Wei X, Chang WJ, Liu Y, Wang ZQ. 2008.** Anatomical traits associated
 931 with absorption and mycorrhizal colonization are linked to root branch order in twenty-
 932 three Chinese temperate tree species. *New Phytologist* **180**: 673-683.
- 933 **Hacke UG, Sperry JS, Ewers BE, Ellsworth DS, Schäfer KVR, Oren R. 2000.** Influence of
 934 soil porosity on water use in *Pinus taeda*. *Oecologia* **124**: 495–505.
- 935 **Hallgren WS, Pitman AJ. 2000.** The uncertainty in simulations by a Global Biome Model
 936 (BIOME3) to alternative parameter values. *Global Change Biology* **6**: 483-495.
- 937 **Hanson PJ, Amthor JS, Wullschleger SD et al. 2004.** Oak forest carbon and water simulations:
 938 model intercomparisons and evaluations against independent data. *Ecological Monographs*
 939 **74(3)**: 443-489.
- 940 **Hawkins BJ, Robbins S, Porter RB. 2014.** Nitrogen uptake over entire root systems of tree
 941 seedlings. *Tree Physiology* **34**: 334–342.
- 942 **Herron PM, Gage DJ, Cardon ZG. 2010.** Micro-scale water potential gradients visualized in
 943 soil around plant root tips using microbiosensors. *Plant Cell and Environment* **33**: 199-210.
- 944 **Hillel D, Van Beek CGEM, Talpaz H. 1975.** A microscopic model of soil water uptake and salt
 945 movement to plants. *Soil Science* **120**: 385-399.
- 946 **Hinsinger P, Brauman A, Devau N, Gérard F, Jourdan C, Laclau JP, Le Cadre E, Jailard**
 947 **B, Plassard C. 2011.** Acquisition of phosphorus and other poorly mobile nutrients by
 948 roots. Where do plant nutrition models fail? *Plant and Soil* **348**: 29-61.

- 949 **Ho MD, McCannon BC, Lynch JP. 2004.** Optimization modeling of plant root architecture for
950 water and phosphorus acquisition. *Journal of Theoretical Biology* **226**: 331-340.
- 951 **Hodge A. 2004.** The plastic plant: root responses to heterogeneous supplies of nutrients. *New*
952 *Phytologist* **162**: 9-24.
- 953 **Hopmans JW, Bristow KL. 2002.** Current capabilities and future needs of root water and
954 nutrient uptake modeling. *Advances in Agronomy* **77**: 103-183.
- 955 **Iversen CM. 2010.** Digging deeper: Fine-root responses to rising atmospheric CO₂
956 concentration in forested ecosystems. *New Phytologist* **186**: 346-357.
- 957 **Iversen CM. 2014.** Using root form to improve our understanding of root function. *New*
958 *Phytologist* **203**: 707-709
- 959 **Iversen CM, Murphy MT, Allen MF, Childs J, Eissenstat DM, Lilleskov EA, Sarjala TM,**
960 **Sloan VL, Sullivan PF. 2012.** Advancing the use of minirhizotrons in wetlands. *Plant and*
961 *Soil* **352**: 23–39.
- 962 **Jackson RB, Manwaring JH, Caldwell MM. 1990.** Rapid physiological adjustment of roots to
963 localized soil enrichment. *Nature* **344**: 58-60.
- 964 **Jackson RB, Canadell J, Ehleringer JR, et al. 1996.** A global analysis of root distributions for
965 terrestrial biomes. *Oecologia* **108**: 389-411.
- 966 **Jackson, RB, HA Mooney, E-D Schulze. 1997.** A global budget for fine root biomass, surface
967 area, and nutrient contents. *Proceedings of the National Academy of Sciences* **94**:7362-
968 7366.
- 969 **Janott M, Gayler S, Gessler A, Javaux M, Klier C, Priesack E. 2011.** A one-dimensional
970 model of water flow in soil-plant systems based on plant architecture. *Plant and Soil* **341**:
971 233-256.
- 972 **Javaux M, Schroder T, Vanderborght J, Vereecken H. 2008.** Use of a three-dimensional
973 detailed modeling approach for predicting root water uptake. *Vadose Zone Journal* **7**:
974 1079-1088.
- 975 **Javot H, Maurel C. 2002.** The role of aquaporins in root water uptake. *Annals of Botany* **90**:
976 301-313.

- 977 **Johnson DM, Sherrard ME, Domec J-C, Jackson RB.** 2014. Role of aquaporin activity in
 978 regulating deep and shallow root hydraulic conductance during extreme drought. *Trees* (in
 979 press)
- 980 **Jourdan C, Rey H.** 1997. Modeling and simulation of the architecture and development of the
 981 oil-palm (*Elaeis guineensis* Jacq.) root system. *Plant and Soil* **190**: 217-233.
- 982 **Kattge J, S Díaz, S Lavorel, IC Prentice et al.** 2011. TRY – a global database of plant traits.
 983 *Global Change Biology* 17:2905–2935.
- 984 **Katul, G. G., R. Oren, S. Manzoni, C. Higgins, and M. B. Parlange.** 2012.
 985 Evapotranspiration: A process driving mass transport and energy exchange in the soil-
 986 plant-atmosphere-climate system. *Reviews of Geophysics* **50**:RG3002,
 987 doi:10.1029/2011RG000366.
- 988 **Kleidon A, Heimann M.** 1998. Optimised rooting depth and its impacts on the simulated
 989 climate of an atmospheric general circulation model. *Geophysical Research Letters* **25**:
 990 345-348.
- 991 **Kleidon A, Heimann M.** 2000. Assessing the role of deep rooted vegetation in the climate
 992 system with model simulations: mechanism, comparison to observations and implications
 993 for Amazonian deforestation. *Climate Dynamics* **16**: 183-199.
- 994 **Kong D, Ma C, Zhang Q, Li L, Chen X, Zeng H, Guo D.** 2014. Leading dimensions in
 995 absorptive root trait variation across 96 subtropical forest species.
 996 *New Phytologist* **203**: 863–872.
- 997 **Koven CD, Riley WJ, Subin ZM, Tang JY, Torn MS, Collins WD, Bonan GB, Lawrence**
 998 **DM, Swenson SC.** 2013. The effect of vertically resolved soil biogeochemistry and
 999 alternate soil C and N models on C dynamics of CLM4. *Biogeosciences* **10**: 7109-7131.
- 1000 **Lambers H, Raven JA, Shaver GR, Smith SE.** 2008. Plant nutrient-acquisition strategies
 1001 change with soil age. *Trends in Ecology and Evolution* **23**: 95-103.
- 1002 **Law BE, Turner D, Lefsky M, Campbell J, Guzy M, Sun O, Van Tuyl S, Cohen W.** 2006.
 1003 Carbon fluxes across regions: Observational constraints at multiple scales. In J. Wu, B.
 1004 Jones, H. Li, O. Loucks, eds. *Scaling and Uncertainty Analysis in Ecology: Methods and*
 1005 *Applications*. Springer, USA. Pages 167-190.

- 1006 **Lee J-E, Oliveira RS, Dawson TE, Fung I. 2005.** Root functioning modifies seasonal climate.
1007 *Proceedings of the National Academy of Sciences, USA* **102**: 17576-17581.
- 1008 **Lehto T, Zwiazek JJ. 2011.** Ectomycorrhizas and water relations of trees: a review. *Mycorrhiza*
1009 **21**: 71-90.
- 1010 **Lindenmair J, Matzner E, Zimmermann R. 2004.** The role of woody roots in water uptake of
1011 mature spruce, beech, and oak trees. In: Matzner, E. (Ed.), *Biogeochemistry of Forested*
1012 *Catchments in a Changing Environment—a German Case Study*. Springer-Verlag, pp. 279–
1013 290.
- 1014 **Loew A, van Bodegom PM, Widlowski J-L, Otto J, Quaife T, Pinty B, Raddatz T. 2013.** Do
1015 we (need to) care about canopy radiation schemes in DGVMs? An evaluation and
1016 assessment study. *Biogeosciences Discussions* **10**: 16551-16613.
- 1017 **Lucash MS, Eissenstat DM, Joslin JD, McFarlane KJ, Yanai RD. 2007.** Estimating nutrient
1018 uptake by mature tree roots under field conditions: challenges and opportunities. *Trees* **21**:
1019 593-603.
- 1020 **Lynch JP, Nielsen KL, Davis RD, Jabllokow AG. 1997.** SimRoot: Modeling and visualization
1021 of root systems. *Plant and Soil* **188**: 139-151.
- 1022 **Manoli G, Bonetti S, Domec J-C, Putti M, Katul G, Marani M. 2014.** Tree root systems
1023 competing for soil moisture in a 3D soil–plant model. *Advances in Water Resources* **66**:
1024 32-42.
- 1025 **Manzoni S, Vico G, Porporato A, Katul G. 2013.** Biological constraints on water transport in
1026 the soil-plant-atmosphere system. *Advances in Water Resources* **51**: 292-304.
- 1027 **Matimati I, Verboom GA, Cramer MD. 2014.** Do hydraulic redistribution and nocturnal
1028 transpiration facilitate nutrient acquisition in *Aspalathus linearis*? *Oecologia* **175**: 1129-
1029 1142.
- 1030 **Matamala R, Stover D B. 2013.** Introduction to a Virtual Special Issue: modeling the hidden
1031 half - the root of our problem. *New Phytologist* **200**: 939-942.
- 1032 **Matzner SL, Richards JH. 1996.** Sagebrush (*Artemisia tridentate* Nutt.) roots maintain nutrient
1033 uptake capacity under water stress. *Journal of Experimental Botany* **47**: 1045-1056.

- 1034 **Maurel C, Verdoucq L, Luu DT, Santoni V. 2008.** Plant aquaporins: membrane channels with
1035 multiple integrated functions. *Annual Review of Plant Biology* **59**:595-624.
- 1036 **May RM. 2004.** Uses and abuses of mathematics in biology. *Science* **303**: 790-793.
- 1037 **McCormack ML, DM Eissenstat, AM Prasad, EAH Smithwick. 2013.** Regional scale
1038 patterns of fine root lifespan and turnover under current and future climate. *Global Change*
1039 *Biology* **19**: 1697-1708.
- 1040 **McCormack, ML, Adams, TS, Smithwick, EAH, Eissenstat, DM. 2012.** Predicting fine root
1041 lifespan from plant functional traits in temperate trees. *New Phytologist* **195**: 823–831. doi:
1042 10.1111/j.1469-8137.2012.04198.x
- 1043 **McMurtrie RE, Dewar RC, Medlyn BE, Jeffreys MP. 2000.** Effects of elevated CO₂ on forest
1044 growth and carbon storage: a modeling analysis of the consequences of changes in litter
1045 quality/quantity and root exudation. *Plant and Soil* **224**: 135-152.
- 1046 **McMurtrie RE, Dewar RC. 2013.** New insights into carbon allocation by trees from the
1047 hypothesis that annual wood production is maximised. *New Phytologist* **199(4)**: 981-990.
1048 DOI: 10.1111/nph.12344.
- 1049 **McMurtrie RE, Iversen CM, Dewar RC, Medlyn BE, Nasholm T, Pepper DA, Norby RJ.**
1050 **2012.** Plant root distributions and N uptake predicted by a hypothesis of optimal root
1051 foraging. *Ecology and Evolution* **2**: 1235-1250.
- 1052 **Meinzer OE. 1927.** Plants as indicators of ground water. USGS Water-Supply Paper 577.
1053 Washington, DC. 95p.
- 1054 **Meinzer FC. 2002.** Co-ordination of vapour and liquid phase water transport properties in
1055 plants. *Plant, Cell and Environment* **25**: 265-274.
- 1056 **Mercado LM, Huntingford C, Gash JHC, Cox PM, Jogireddy V. 2007.** Improving the
1057 representation of radiation interception and photosynthesis for climate model applications.
1058 *Tellus Series B-Chemical and Physical Meteorology* **59**: 553-565.
- 1059 **Mills R, Lu C, Lichtner PC, Hammond G. 2007.** Simulating Subsurface Flow and Transport
1060 on Ultrascale Computers using PFLOTRAN. *Journal of Physics Conference Series* **78**:
1061 012051 doi:10.1088/1742-6596/78/1/012051.

- 1062 **Molz FJ. 1981.** Models of water transport in the soil-plant system: A review. *Water Resources*
1063 *Research* **17**: 1245-1260.
- 1064 **Molz FJ, Remson I. 1970.** Extraction term models of soil moisture use by transpiring plants.
1065 *Water Resources Research* **6(5)**: 1346–1356. doi:10.1029/WR006i005p01346.
- 1066 **Nadezhdina N, Cermak J. 2003.** Instrumental methods for studies of structure and function of
1067 root systems of large trees. *Journal of Experimental Botany* **54**: 1511-1521.
- 1068 **Neumann RB, Cardon ZG. 2012.** The magnitude of hydraulic redistribution by plant roots: a
1069 review and synthesis of empirical and modeling studies. *New Phytologist* **194**: 337-352.
- 1070 **Nielsen KL, Lynch JP, Jabllokow AG, Curtis PS. 1994.** Carbon cost of root systems: an
1071 architectural approach. *Plant and Soil* **165**: 161-169.
- 1072 **Norby RJ, Jackson RB. 2000.** Root dynamics and global change: seeking an ecosystem
1073 perspective. *New Phytologist* **147**: 1-12.
- 1074 **Nye PH. 1966.** The effect of nutrient intensity and buffering power of a soil, and the absorbing
1075 power, size and root-hairs of a root, on nutrient absorption by diffusion. *Plant and Soil* **25**:
1076 81-105.
- 1077 **Nye PH, Marriot FHC. 1969.** A theoretical study of the distribution of substances around roots
1078 resulting from simultaneous diffusion and mass flow. *Plant and Soil* **30**: 459-472.
- 1079 **Oleson KW, Lawrence DM, Bonan GB et al. 2010.** Technical description of version 4.0 of the
1080 Community Land Model (CLM), NCAR Tech. Note NCAR/TN-478+STR, 257 pp., Natl.
1081 Cent. for Atmos. Res., Boulder, Colo.
- 1082 **Oleson K, Lawrence DM, Bonan GB et al. 2013.** Technical description of version 4.5 of the
1083 Community Land Model (CLM). NCAR Technical Note NCAR/TN-503+STR, 420 pp,
1084 DOI: 10.5065/D6RR1W7M.
- 1085 **Ostle NJ, Smith P, Fisher R, Woodward FI, Fisher JB, Smith JU, Galbraith D, Levy P, Meir**
1086 **P, McNamara NP, Bardgett RD. 2009.** Integrating plant-soil interactions into global
1087 carbon cycle models. *Journal of Ecology* **97**: 851-863.
- 1088 **Pagès L, Jordan MO, Picard D. 1989.** A simulation model of the three-dimensional
1089 architecture of the maize root system. *Plant and Soil* **119**: 147-154.

- 1090 **Pagès L, Vercambre G, Drouet J-L, Lecompte F, Collet C, Le Bot J. 2004.** Root Typ: a
1091 generic model to depict and analyse the root system architecture. *Plant and Soil* **258**: 103-
1092 119.
- 1093 **Pan Y, McGuire AD, Melillo JM, Kicklighter DW, Sitch S, Prentice IC. 2002.** A
1094 biogeochemistry-based dynamic vegetation model and its application along a moisture
1095 gradient in the continental United States. *Journal of Vegetation Science* **13**: 369-382.
- 1096 **Parton WJ, Stewart JWB, Cole CV. 1988.** Dynamics of c, n, p and s in grassland soils - a
1097 model. *Biogeochemistry* **5**: 109-131.
- 1098 **Peters DPC, Loescher HW, SanClements MD, Havstad KM. 2014.** Taking the pulse of a
1099 continent: expanding site-based research infrastructure for regional- to continental-scale
1100 ecology. *Ecosphere* **5(3)**: 29.
- 1101 **Pierret A, Moran CJ, Doussan C. 2005.** Conventional detection methodology is limiting our
1102 ability to understand the roles and functions of fine roots. *New Phytologist* **166**:967-980.
- 1103 **Pitman AJ. 2003.** The evolution of, and revolution in, land surface schemes designed for climate
1104 models. *International Journal of Climatology* **23**: 479-510.
- 1105 **Pohl M, Stroude R, Buttler A, Rixen C. 2011.** Functional traits and root morphology of alpine
1106 plants. *Annals of Botany* **108**: 537-545.
- 1107 **Postma JA, Lynch JP. 2011a.** Root cortical aerenchyma enhances the acquisition and utilization
1108 of N, phosphorus, and potassium in *Zea mays* L. *Plant Physiology* **156**: 1190-1201.
- 1109 **Postma JA, Lynch JP. 2011b.** Theoretical evidence for the functional benefit of root cortical
1110 aerenchyma in soils with low phosphorus availability. *Annals of Botany* **107**: 829-841.
- 1111 **Pritchard SG, Strand AE 2008.** Can you believe what you see? Reconciling minirhizotron and
1112 isotopically derived estimates of fine root longevity. *New Phytologist* **177**: 287-291.
- 1113 **Qu Y, Duffy CJ. 2007.** A semi-discrete finite volume formulation for multiprocess watershed
1114 simulation. *Water Resources Research* **43**: W08419.
- 1115 **Raats PAC. 2007.** Uptake of water from soils by plant roots. *Transport in Porous Media* **68**: 5-
1116 28.
- 1117 **Raich JW, Rastetter EB, Melillo JM, et al. 1991.** Potential net primary productivity in South
1118 America: application of a global model. *Ecological Applications* **1**: 399-429.

- 1119 **Rastetter EB, Ryan MG, Shaver GR, Melillo JM, Nadelhoffer KJ, Hobbie JE, Aber JD.**
1120 **1991.** A general biogeochemical model describing the responses of the C-cycle and N-
1121 cycle in terrestrial ecosystems to changes in CO₂, climate, and N-deposition. *Tree*
1122 *Physiology* **9**: 101-126.
- 1123 **Read DJ, Boyd R. 1986.** Water relations of mycorrhizal fungi and their host plants. In: Ayres
1124 PG, Boddy L, eds. *Water, fungi and plants*. Cambridge, UK: Press Syndicate and
1125 University of Cambridge, 287-303.
- 1126 **Rengel Z. 1993.** Mechanistic simulation models of nutrient uptake: A review. *Plant and Soil*
1127 **152**: 161-173.
- 1128 **Rewald B, Ephrath JE, Rachmilevitch S. 2011.** A root is a root is a root? Water uptake rates of
1129 citrus root orders. *Plant Cell and Environment* **34**: 33-42.
- 1130 **Rihani JF, Maxwell RM, Chow FK. 2010.** Coupling groundwater and land surface processes:
1131 Idealized simulations to identify effects of terrain and subsurface heterogeneity on land
1132 surface energy fluxes. *Water Resources Research* **46**: W12 523.
- 1133 **Riley, WJ, Gaudinski, JB, Torn, MS, Dawson, TE, Joslin, JD, Majdi H. 2009.** Fine-root
1134 mortality rates in a temperate forest: Estimates using radiocarbon data and numerical
1135 modeling, *New Phytologist* **184**: 387–398, doi:10.1111/j.1469-8137.2009.02980.x.
- 1136 **Running SW, Coughlan JC. 1988.** A general-model of forest ecosystem processes for regional
1137 applications .1. Hydrologic balance, canopy gas-exchange and primary production
1138 processes. *Ecological Modeling* **42**: 125-154.
- 1139 **Schenk HJ, Jackson RB. 2002.** The global biogeography of roots. *Ecological Monographs* **72**:
1140 311-328.
- 1141 **Schenk HJ. 2005.** Vertical vegetation structure below ground: Scaling from root to globe.
1142 *Progress in Botany* **66**: 341-373.
- 1143 **Schenk HJ. 2008.** The shallowest possible water extraction profile: A null model for global root
1144 distributions. *Vadose Zone Journal* **7**: 1119-1124.
- 1145 **Schneider CL, Attinger S, Delfs JO, Hildebrandt A. 2010.** Implementing small scale processes
1146 at the soil-plant interface - the role of root architectures for calculating root water uptake
1147 profiles. *Hydrology and Earth Systems Science* **14**: 279-289.

- 1148 **Schnepf A, Leitner D, Klepsch S. 2012.** Modeling phosphorus uptake by a growing and
1149 exuding root system. *Vadose Zone Journal* **11**. doi:10.2136/vzj2012.0001.
- 1150 **Scholz FG, Bucci SJ, Goldstein G, Meinzer FC, Franco AC, Miralles-Wilhelm F. 2007.**
1151 Biophysical properties and functional significance of stem water storage tissues in
1152 Neotropical savanna trees. *Plant, Cell, and Environment* **30**: 236-248
- 1153 **Schroder T, Javaux M, Vanderborght J, Korfgan B, Vereecken H. 2008.** Effect of local soil
1154 hydraulic conductivity drop using a 3-D root water uptake model. *Vadose Zone Journal* **7**:
1155 1089-1098.
- 1156 **Schroder T, Javaux M, Vanderborght J, Korfgan B, Vereecken H. 2009.** Implementation of a
1157 microscopic soil-root hydraulic conductivity drop function in a 3-D soil-root architecture
1158 water transfer model. *Vadose Zone Journal* **8**: 783-792.
- 1159 **Schulze, ED. 2013.** Large-scale biogeochemical research with particular reference to forest
1160 ecosystems, an overview. *Forest Ecology and Management* (in press)
1161 <http://dx.doi.org/10.1016/j.foreco.2013.07.054>
- 1162 **Schymanski, SJ, M Sivapalan, ML Roderick, J Beringer, LB Hutley. 2008.** An optimality-
1163 based model of the coupled soil moisture and root dynamics. *Hydrology and Earth System*
1164 *Sciences* **12**: 913-932.
- 1165 **Segal E, Kushnir T, Mualem Y, Shani U. 2008.** Water uptake and hydraulics of the root hair
1166 rhizosphere. *Vadose Zone Journal* **7**: 1027-1034.
- 1167 **Shi Y, Davis KJ, Duffy CJ, Xuan Y. 2013.** Development of a coupled land surface hydrologic
1168 model and evaluation at a critical zone observatory. *Journal of Hydrometeorology* **14**:
1169 1401-1420.
- 1170 **Siemens JA, Zwiazek JJ. 2004.** Changes in root water flow properties of solution culture-grown
1171 trembling aspen (*Populus tremuloides*) seedlings under different intensities of water-deficit
1172 stress. *Physiologia Plantarum* **121**: 44-49.
- 1173 **Šimůnek J, Hopmans JW. 2009.** Modeling compensated root water and nutrient uptake.
1174 *Ecological Modelling* **220**: 505–521.

- 1175 **Sivandran G, Bras RL. 2013.** Dynamic root distribution in ecohydrological modeling: A case
1176 study at Walnut Gulch Experimental Watershed., *Water Resources Research* **49**: 3292-
1177 3305.
- 1178 **Skaggs TH, van Genuchten MT, Shouse PJ, Poss JA. 2006.** Macroscopic approaches to root
1179 water uptake as a function of water and salinity stress. *Agricultural Water Management* **86**:
1180 140-149.
- 1181 **Smith B, Wårlind D, Arneeth A, Hickler T, Leadley P, Siltberg J, Zaehle S. 2013.** Implications
1182 of incorporating N cycling and N limitations on primary production in an individual-based
1183 dynamic vegetation model. *Biogeosciences Discussions* **10**: 18613-18685.
- 1184 **Somma F, Hopmans JW, Clausnitzer V. 1998.** Transient three-dimensional modeling of soil
1185 water and solute transport with simultaneous root growth, root water and nutrient uptake.
1186 *Plant and Soil* **202**: 281-293.
- 1187 **Spek LY. 1997.** Generation and visualization of root-like structures in a three-dimensional space.
1188 *Plant and Soil* **197**: 9-18.
- 1189 **Sperry JS, Adler FR, Campbell GS, Comstock JP. 1998.** Limitation of plant water use by
1190 rhizosphere and xylem conductance: results from a model. *Plant, Cell and Environment*
1191 **21**: 347-59.
- 1192 **Stedle E. 2000.** Water uptake by roots: effects of water deficit. *Journal of Experimental Botany*
1193 **51**: 1531-1542.
- 1194 **Stedle E. 2001.** The cohesion-tension mechanism and the acquisition of water by plant roots.
1195 *Annual Review of Plant Physiology and Plant Molecular Biology* **52**: 847-875.
- 1196 **Thaler P, Pagès L. 1998.** Modeling the influence of assimilate availability on root growth and
1197 architecture. *Plant and Soil* **201**: 307-320.
- 1198 **Thomas RQ, Bonan GB, Goodale CL. 2013.** Insights into mechanisms governing forest carbon
1199 response to N deposition: a model-data comparison using observed responses to N
1200 addition. *Biogeosciences* **10**: 3869-3887.
- 1201 **Thornton PE, Lamarque J-F, Rosenbloom NA, Mahowald NM. 2007.** Influence of carbon-N
1202 cycle coupling on land model response to CO₂ fertilization and climate variability. *Global*
1203 *Biogeochemical Cycles* **21**: GB4018.

- 1204 **Valenzuela-Estrada LR, Vera-Caraballo V, Ruth LE, Eissenstat DM. 2008.** Root anatomy,
1205 morphology and longevity among root orders in *Vaccinium corymbosum* (Ericaceae).
1206 *American Journal of Botany* **95**: 1506-1514.
- 1207 **Van Rees KCJ, Comerford NB. 1990.** The role of woody roots of slash pine seedlings in water
1208 and potassium absorption. *Canadian Journal of Forest Research* **20**:1183-1191.
- 1209 **Vandeleur RK, Sullivan W, Athman A, Jordans C, Gilliham M, Kaiser BN, Tyerman SD.**
1210 **2014.** Rapid shoot-to-root signalling regulates root hydraulic conductance via aquaporins.
1211 *Plant, Cell and Environment* **37**: 520-538.
- 1212 **Vargas R, Allen MF. 2008.** Dynamics of fine root, fungal rhizomorphs, and soil respiration in a
1213 mixed temperate forest: integrating sensors and observations. *Vadose Zone Journal* **7**:
1214 1055-1064.
- 1215 **Verheijen, LM, Brovkin, V, Aerts, R, Bönisch, G, Cornelissen, JHC, Kattge, J, Reich, PB,**
1216 **Wright, IJ, van Bodegom, PM. 2013.** Impacts of trait variation through observed trait–
1217 climate relationships on performance of an Earth system model: a conceptual analysis,
1218 *Biogeosciences* **10**: 5497-5515. doi:10.5194/bg-10-5497-2013.
- 1219 **Verhoef A, Egea G. 2014.** Modeling plant transpiration under limited soil water: Comparison of
1220 different plant and soil hydraulic parameterizations and preliminary implications for their
1221 use in land surface models. *Agricultural and Forest Meteorology* **191**: 22–32.
- 1222 **Vermeer, JEM, von Wangenheim D, Barberon M, Lee Y, Ernst HK, Stelzer AM, Geldner N.**
1223 **2014.** A spatial accommodation by neighboring cells is required for organ initiation in
1224 *Arabidopsis*. *Science* **343**: 178-183.
- 1225 **Vrugt JA, van Wijk MT, Hopmans JW, Simunek J. 2001.** One-, two-, and three-dimensional
1226 root water uptake functions for transient modeling. *Water Resources Research* **37**: 2457-
1227 2470.
- 1228 **Walker AP, Hanson PJ, De Kauwe MG, Medlyn BE, Zaehle S, Asao S, Dietze MC, Hickler**
1229 **T, Huntingford C, Iversen C, et al. 2014.** Model-experiment synthesis at two temperate
1230 forest free-air CO₂ enrichment experiments. *Journal of Geophysical Research -*
1231 *Biogeosciences* (in press).

- 1232 **Wang, D, Xu Y, Thornton PE, King AW, Gu L, Steed C. 2014.** A functional testing platform
1233 for the community land model, environmental modeling and software. *Environmental*
1234 *Modeling and Software* **55**: 25–31.
- 1235 **Wang G, Alo C, Mei R, Sun S. 2011.** Droughts, hydraulic redistribution, and their impact on
1236 vegetation composition in the Amazon forest. *Plant Ecology* **212**: 663–673.
- 1237 **Wang S, Grant RF, Verseghy DL, Black TA. 2002.** Modelling carbon-coupled energy and
1238 water dynamics of a boreal aspen forest in a general circulation model land surface
1239 scheme. *International Journal of Climatology* **22**: 1249–1265.
- 1240 **Wang YP, Law RM, Pak B. 2010.** A global model of carbon, N and phosphorus cycles for the
1241 terrestrial biosphere. *Biogeosciences* **7**: 2261–2282.
- 1242 **Warren JM, Brooks JR, Dragila MI, Meinzer FC. 2011.** In situ separation of root hydraulic
1243 redistribution of soil water from liquid and vapor transport. *Oecologia* **166**: 899–911.
- 1244 **Warren JM, Meinzer FC, Brooks JR, Domec JC. 2005.** Vertical stratification of soil water
1245 storage and release dynamics in Pacific Northwest coniferous forests. *Agricultural and*
1246 *Forest Meteorology* **130**: 39–58.
- 1247 **Warren JM, Bilheux H, Kang M, Voisin S, Cheng C, Horita J, Perfect E. 2013.** Neutron
1248 imaging reveals internal plant water dynamics. *Plant and Soil* **366**: 683–693.
- 1249 **Wilkinson S, Davies WJ. 2002.** ABA-based chemical signaling: the co-ordination of responses
1250 to stress in plants. *Plant, Cell and Environment* **25**: 195–210.
- 1251 **Williams M, Rastetter EB, Fernandes DN, et al. 1996.** Modeling the soil-plant-atmosphere
1252 continuum in a *Quercus-Acer* stand at Harvard forest: the regulation of stomatal
1253 conductance by light, nitrogen and soil-plant hydraulic properties. *Plant, Cell and*
1254 *Environment* **19**: 911–927.
- 1255 **Woodward FI, Lomas MR. 2004.** Vegetation dynamics - simulating responses to climatic
1256 change, *Biological Reviews* **79**: 643–670.
- 1257 **Woodward FI, Osborne CP. 2000.** The representation of root processes in models addressing
1258 the responses of vegetation to global change. *New Phytologist* **147**: 223–232.
- 1259 **Wu L, McGechan MB. 1998.** Simulation of biomass, carbon and N accumulation in grass to
1260 link with a soil N dynamics model. *Grass Forage Science* **53**: 233–249.

- 1261 **Wu L, McGechan MB, McRoberts N, Baddeley JA, Watson CA. 2007.** SPACSYS:
1262 integration of a 3D root architecture component to carbon, N, and water cycling - model
1263 description. *Ecological Modeling* **200**: 343-359.
- 1264 **Wullschleger SD, Epstein HE, Box EO, Euskirchen ES, Goswami S, Iversen CM, Kattge J,**
1265 **Norby RJ, van Bodegom PM, Xu X. 2014.** Plant functional types in Earth System
1266 Models: Past experiences and future directions for application of dynamic vegetation
1267 models in high-latitude ecosystems. *Annals of Botany* **114**: 1-16.
- 1268 **Young IM. 1998.** Biophysical interactions at the root-soil interface: a review. *Journal of*
1269 *Agricultural Science, Cambridge.* **130**: 1-7.
- 1270 **Zaehle S, Friend AD. 2010.** Carbon and N cycle dynamics in the O-CN land surface model: 1.
1271 Model description, site-scale evaluation, and sensitivity to parameter estimates. *Global*
1272 *Biogeochemical Cycles* **24**: GB1005.
- 1273 **Zaehle S, Medlyn BE, De Kauwe MG, Walker AP, Dietze MC, Hickler T, Luo Y, Wang Y-P,**
1274 **El-Masri B, Thornton P, et al. 2014.** Evaluation of 11 terrestrial carbon–nitrogen cycle
1275 models against observations from two temperate Free-Air CO₂ Enrichment studies. *New*
1276 *Phytologist* **202**: 803–822.
- 1277 **Zeppel MJB, Lewis JD, Chaszar B, Smith RA, Medlyn BE, Huxman TE, Tissue DT. 2012.**
1278 Nocturnal stomatal conductance responses to rising [CO₂], temperature and drought. *New*
1279 *Phytologist* **193**: 929–938.
- 1280

1281 **Figure Titles**

1282

1283 **Figure 1.** Diagram of the structural and functional characteristics of fine roots of plant root
1284 systems, and their interaction with the soil rhizosphere. Developing fine roots contain zones of
1285 active growth and function and zones where changes in anatomical tissue reduces root functions
1286 such as water or nutrient uptake. Water and solutes can move passively through the apoplast of
1287 the epidermis, cortex and young developing endodermis to the central vascular tissue. As the root
1288 tissue matures endodermal cell walls become suberized, at which point water and nutrients
1289 uptake into the symplast is regulated by passive or active transport proteins, such as aquaporins
1290 (water) or ion-pumps (mineral nutrients). Functionality of fine roots varies with characteristic
1291 morphological traits that are specific to species, and that respond to soil biotic and abiotic
1292 signals, such as mycorrhizae or soil drying.. In this diagram functions associated with nutrient
1293 uptake are presented in orange text, water transport in blue text, and carbon transport in green
1294 text.

1295

1296 **Figure 2.** Advanced techniques illustrate novel insight into root structure and dynamic root
1297 processes, such as (a) Ericaceous shrub roots and associated mycorrhizal hyphae and (b) a fungal
1298 rhizomorph from an automated minirhizotron system deployed in a peatbog (scale $\sim 2.5 \times 3$ mm);
1299 (c) scanning electron micrograph of ~ 30 - 50 μm -long root hairs of *Quercus rubra*; (d-g) neutron
1300 imaging time-series of water uptake and internal transport (orange colors) through corn seedlings
1301 over ~ 12 hours following a pulse of water below the roots (blue). Such data can be used to
1302 validate model simulations of root structure, production, turnover and water uptake.

1303

1304 **Figure 3.** Root, whole plant, and terrestrial biosphere models (TBMs) in relation to spatial and
1305 temporal scales at which they operate. Mechanistic root processes are readily modeled for single
1306 roots, but process-based knowledge is dramatically lost for higher-order models, resulting in
1307 more static and less complex representation as spatial scale increases. Landscape-level bulk root
1308 distribution, water and nutrient uptake are estimated and not dynamic in most TBMs. Root traits
1309 can provide a framework for scaling dynamic root functions (such as fine root proliferation, loss

1310 of root conductivity, or hydraulic redistribution) into TBMs to improve model veracity – a
1311 pathway indicated by the large arrow.

1312

1313 **Figure 4.** Key root structural and functional attributes and their inclusion in several well-known
1314 ecosystem and terrestrial biosphere models (TBMs) – filled circles represent model inclusion.
1315 Dynamic root functions such as Michaelis-Menten (M-M) nutrient uptake kinetics, hydraulic
1316 redistribution of water (HR) and downregulation due to low oxygen (Anoxia) are rarely included
1317 in the models. Other functions such as water uptake are widely represented when linked
1318 specifically to root depth, but rarely consider actual root biomass. Model references as in Tables
1319 2, 3.

1320

1321 **Figure 5.** Framework for assessment of root data, and its importance in scaling ecosystem
1322 function through root traits for modeling the terrestrial biosphere. (left) Root distribution is the
1323 most common dataset available, and is used in many TBMs to regulate water use (Fig 4).
1324 Improved modeling will include root structural traits (e.g., size, age, order, display, C:N,
1325 mycorrhizal associations), and their associated functions (e.g., water and nutrient uptake, and
1326 carbon release through respiration, exudation and turnover). (right) Model evaluation should first
1327 assess the presence of roots or root functions, including both direct (e.g., water uptake based on
1328 root distribution) and indirect (e.g., nitrogen uptake based on plant demand) functions. Efforts
1329 must be made to understand the role of roots for specific processes at the appropriate spatial and
1330 temporal scales (Fig 3). Key root functions should be prioritized based on current mechanistic
1331 knowledge of root processes and dynamic biotic/abiotic regulation of those processes, as well as
1332 by their relative importance to the model. Addition of new root functionality to a model will
1333 require development of trait databases that can be scaled across landscapes based on species and
1334 plant functional type (PFT) characteristics, soil and environmental conditions.

Table 1. Five individual plant models that represent carbon allocation, root architecture and uptake of water and nutrients.

Model	Allocation	Architecture	Acquisition	Reference
ROOTMAP	Calculates balance between plant demand and the capacity of individual roots to supply soil resources, to drive allocation of assimilates and resultant growth of root tips and branching	Basic attributes affecting growth are elongation rate, branching density, direction, initiation times, and duration of apical non-branching with sensitivities to temperature and soil density	Water uptake is based on a sink term; nitrate uptake is an approximate solution to the convection–dispersion equation using Michaelis-Menten kinetics	Diggle, 1988; Dunbabin et al., 2002; Dunbabin et al., 2003
Root Typ	Allocation to growth occurs at a potential rate for all sinks when sufficient carbohydrate is available; else, reduced growth is determined <i>with</i> or <i>without</i> competing source-sink priorities	Root tips interact with soil temperature, mechanical impedance, and oxygen status to determine root elongation, direction, branching, radial growth, decay, and abscission	Water transfer into and along the root is represented by a set of connected hydraulic axial conductances and radial conductivities distributed within the root system	Pagès et al., 1989; Thaler and Pagès, 1998; Pagès et al., 2004; Doussan et al., 2006
R-SWMS	Root growth is described in three ways; most complex application root growth is a function of dynamic allocation of assimilate to shoot and root (Level 3)	Root axes are generated at defined times; branching and spacing are a function of root age; sensitive to temperature, soil strength, and solute concentration	Water transfer represented by axial and radial conductances as a function of root age and root type; nutrient transport described by convection-diffusion equation	Somma et al., 1998; Javaux et al., 2008; de Willigen et al. 2012
<i>SimRoot</i>	Carbon allocation rules based on a hierarchical binary partitioning method where sink strength, priority, and limits determine the carbon allocated to competing sinks	Spatial patterns determined by types of root branches, branch angles, growth velocities, and sensitivities to temperature, nutrient stress, and carbon availability	Nutrient (N, P, K) uptake is a function of root class, root development, root hair development, and intra-root competition; water uptake not represented in current model	Nielsen et al., 1994; Lynch et al., 1997; Postma and Lynch, 2011a, b
SPACSYS	Roots receive photosynthate with the highest priority; allocation is dependent on plant developmental stage; elongation and volume expansion depend on carbohydrate supply	Root system develops based on elongation rates of various root types, growth direction, branching, and mortality; processes are sensitive to soil temperature, soil strength, and solute concentration	N uptake depends on the concentration of nutrient at the root surface and the kinetics of uptake; water uptake is determined by a localized extraction function modified by soil water potential	Wu and McGechan, 1998; Wu et al., 2007

Table 2. The representation of carbon allocation, root architecture and uptake of water and nutrients in a subset of ecosystem models.

Model	Time step	Allocation		Architecture/ Distribution	Acquisition /Ecosystem Function		
		Carbon	Phenology	By depth	Water uptake	N uptake	Root Turnover & C loss
ECOSYS ¹	Hourly	Functional balance of N, P Demand adjusted so that allocation increases when root storage C:N/C:P > than that required to support new growth	Remaining C from R – MR is available for GR subject to water and N&P status; resistance from soil and root and myco turgor. Allocated to each root by comparative conductance	Controlled by primary root growth, distribution by primary root length and secondary root lengths	$f(\text{root radial and axial resistances, soil water content})$ Uptake (Q) = $(\psi_{\text{shoot}} - \psi_{\text{soil}}) / (\text{sum of radial and axial resistances})$	$f(\text{root N, P})$ diffusion, mass transport, adsorption, microbial immobilization so that uptake = solution concentration at root surface. Demand adjusted so that uptake is inhibited when root storage C:N/C:P > than that required to support new growth	Maintenance respiration (MR, priority): $f(\text{soil temperature, O}_2)$ Growth respiration (GR): $f(\text{water, N, P})$ Nutrient uptake respiration (N_uR): Exudation Turnover: if(MR < M + GR) M&GR = $f(T, O_2)$ status, comparative C conductance, turgor)
G'DAY ²	Daily / Weekly	Fixed fraction of NPP	None	None	Assumed non-limiting; no specific uptake function Updated model version will have two layers with root proportion linked to uptake	Not root-specific: fixed fraction of net soil N mineralization	Respiration: Fixed fraction of GPP; not root-specific Exudation: Fixed fraction of NPP Turnover: Equal to 1.0
SPA ³	30 minutes	Prescribed	None	Maximum root biomass per unit soil volume prescribed; exponential decline in biomass with depth to a prescribed maximum rooting depth. Dmax input parameter as is max root mass in soil volume	$f(\text{root and soil hydraulic resistance, root biomass and distribution, soil water content})$ $E_{\text{max}} = (\psi_{\text{shoot}} - \psi_{\text{soil}}) / (\text{sum of plant resistances})$ capacitance accounted for ψ_{soil} is weighted by root distribution and soil resistance	None	None

Model	Time step	Allocation		Architecture/ Distribution	Acquisition /Ecosystem Function		
TEM ⁴	1 month	none	none	Max rooting depth used to estimate water availability	f (ET demand, soil properties, SWC)	f (soil available N, SWC, C:N energy balance)	f (NPP), above and belowground C loss is single term

¹Grant 1998, ²McMurtrie et al. 2000, ³Williams et al. 1996, ⁴Raich et al. 1991

For Peer Review

Table 3. The representation of carbon allocation, root architecture and uptake of water and nutrients in a subset of terrestrial biosphere models (TBMs) and dynamic global vegetation models.

Model	Time Step	Allocation		Architecture/ Distribution	Acquisition /Ecosystem Function		
		Carbon	Phenology	By depth	Water uptake	N uptake	Root Turnover & C loss
CLM4.0 ¹ CLM4.5 ²	30 minutes	Fixed fraction (1:1 leaf allocation)	Same as leaf	CLM4.0 Double-exponential for water (PFT specific) CLM4.5 Double-exponential for water; exponential for C inputs (PFT specific)	f (plant demand, root distribution, soil matric potential)	If supply > demand, N uptake = demand to meet growth requirements If supply < demand, N uptake = f (soil mineral N, plant demand, microbial demand) (no root dependence)	Linked 1:1 to leaf turnover
CABLE ³	30 minutes	Fixed fraction (varied by phenological phase)	Phased, opposite to leaf phenology	Decreasing proportion with depth	f (plant demand, root proportion, SWC)	f (soil mineral N, plant demand)	Fixed fraction
LM3 ⁴	30 minutes	Functional balance: to maintain root:shoot ratio, root:shoot ratio f (water stress)	Same as leaf			Michaelis-Menten kinetics f (soil mineral N, root mass)	
JULES ⁵	30 minutes	Fixed fraction (1:1 leaf allocation)	Growth: same as leaf Turnover: fixed fraction	Exponential	f (plant demand, root proportion, SWC)	na	Fixed fraction 0.15-0.25 yr ⁻¹
O-CN ⁶	30 minutes to 1 day	Functional balance: to maintain root:shoot ratio, root:shoot ratio f (water or N stress)	balance between allocation and turnover	Decreasing with depth (2 soil layers)	f (plant demand, root proportion, SWC)	Michaelis-Menten kinetics f (soil mineral N, root mass, plant demand, temperature)	f (age) mean turnover rate of 0.7 yr ⁻¹
SDGVM ⁷	1 day	Fixed fraction: 0.0015	If GPP > 0	Fixed proportions	f (plant demand,	f (soil C)	f (age) and self-

Model	Time Step	Allocation		Architecture/ Distribution	Acquisition /Ecosystem Function		
		of labile C pool		through 4 soil layers. 0.5,0.3,0.15,0.05	root proportion, SWC)		thinning mortality
LPJ-GUESS ⁸	1 day	Functional balance: to maintain root:shoot ratio, root:shoot ratio f (water or N stress)	None	Decreasing with depth (2 soil layers)	f (plant demand, root proportion, SWC)	f (soil mineral N, root mass, plant demand, soil T)	Fixed fraction 0.5–0.7 yr ⁻¹
MBL-GEM III ⁹	1 month	Functional balance	Result of allocation	None	na	f (root N content, air T)	Fixed fraction 0.164 yr ⁻¹
DVM-DOS-TEM ¹⁰	1 month	Fixed fraction	Same as leaf	Exponential to max rooting depth	f (plant demand, root proportion, SWC)	f (plant demand, root proportion and mass, root respiration, air T, SWC, available soil N)	f (standing crop, production) 0.25–1 yr ⁻¹

¹Thornton et al. 2007, Oleson et al. 2010, ²Koven et al. 2013, Oleson et al. 2013, ³Wang et al. 2010, ⁴Gerber et al. 2010, ⁵Clark et al. 2011, ⁶Zaehle and Friend 2010, ⁷Woodward and Lomas 2004, ⁸Smith et al. 2013, ⁹Rastetter et al. 1991, ¹⁰Euskirchen et al. 2009

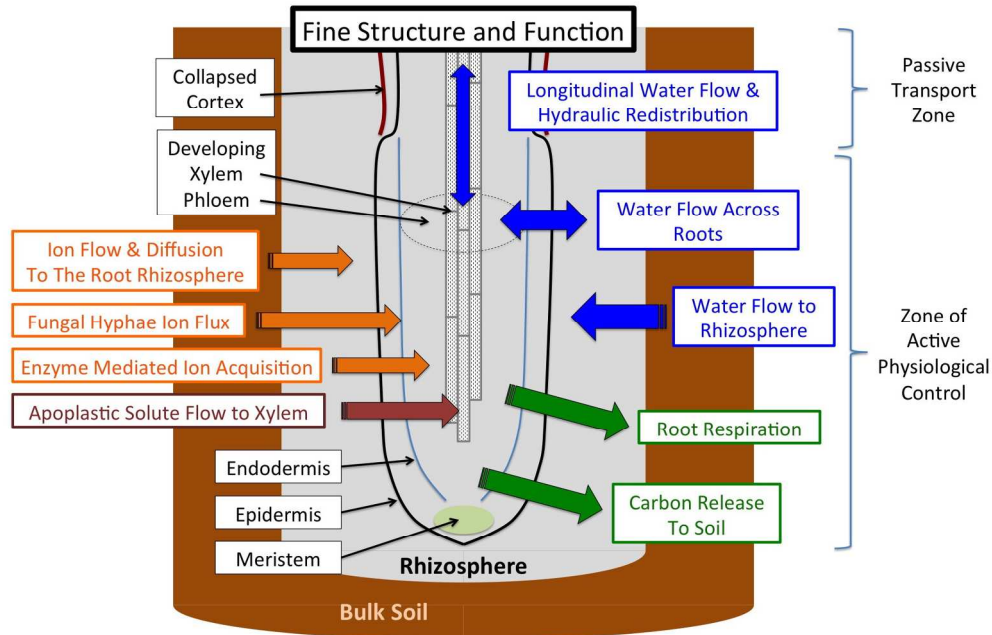


Figure 1. Diagram of the structural and functional characteristics of fine roots of plant root systems, and their interaction with the soil rhizosphere. Developing fine roots contain zones of active growth and function and zones where changes in anatomical tissue reduces root functions such as water or nutrient uptake. Water and solutes can move passively through the apoplast of the epidermis, cortex and young developing endodermis to the central vascular tissue. As the root tissue matures endodermal cell walls become suberized, at which point water and nutrients uptake into the symplast is regulated by passive or active transport proteins, such as aquaporins (water) or ion-pumps (mineral nutrients). Functionality of fine roots varies with characteristic morphological traits that are specific to species, and that respond to soil biotic and abiotic signals, such as mycorrhizae or soil drying.. In this diagram functions associated with nutrient uptake are presented in orange text, water transport in blue text, and carbon transport in green text.

746x477mm (72 x 72 DPI)

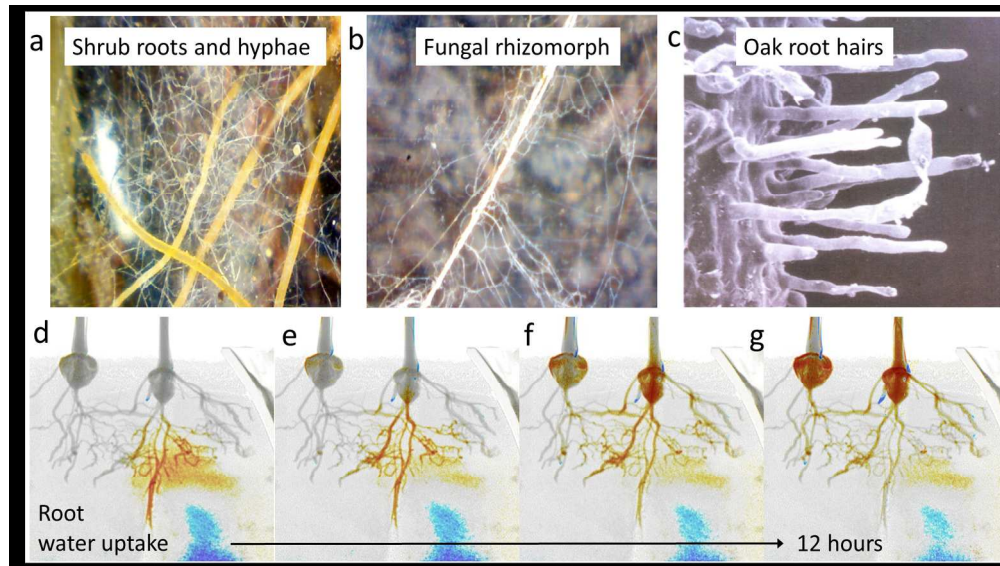


Figure 2. Advanced techniques illustrate novel insight into root structure and dynamic root processes, such as (a) Ericaceous shrub roots and associated mycorrhizal hyphae and (b) a fungal rhizomorph from an automated minirhizotron system deployed in a peatbog (scale $\sim 2.5 \times 3$ mm); (c) scanning electron micrograph of ~ 30 - 50 μm -long root hairs of *Quercus rubra*; (d-g) neutron imaging time-series of water uptake and internal transport (orange colors) through corn seedlings over ~ 12 hours following a pulse of water below the roots (blue). Such data can be used to validate model simulations of root structure, production, turnover and water uptake.

344x195mm (150 x 150 DPI)

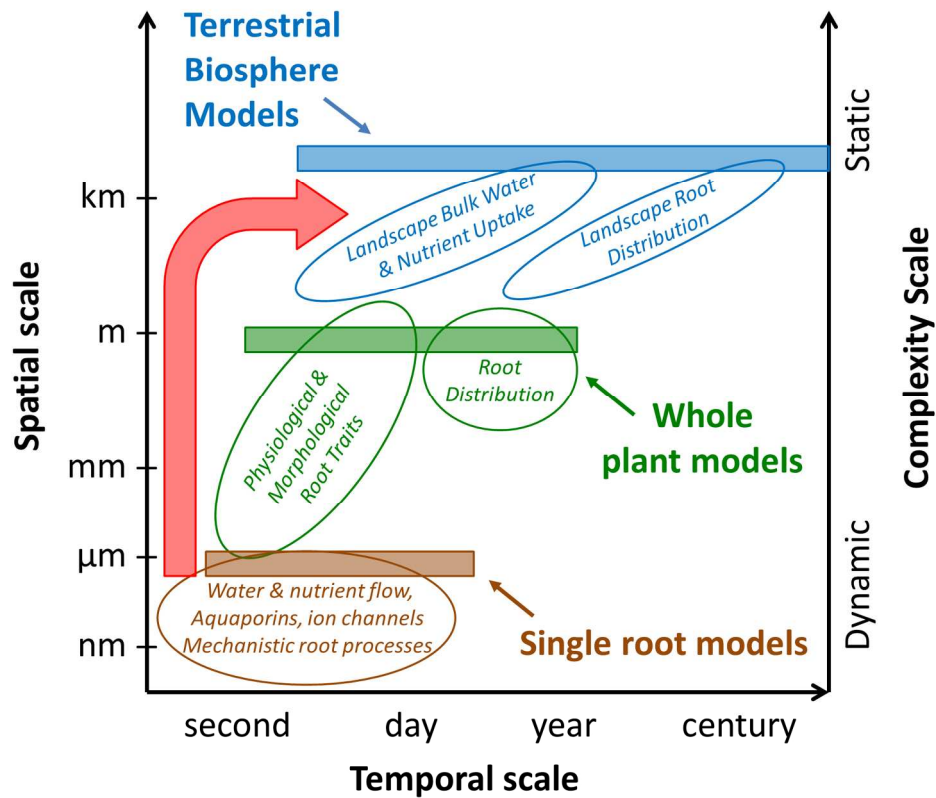


Figure 3. Root, whole plant, and terrestrial biosphere models (TBMs) in relation to spatial and temporal scales at which they operate. Mechanistic root processes are readily modeled for single roots, but process-based knowledge is dramatically lost for higher-order models, resulting in more static and less complex representation as spatial scale increases. Landscape-level bulk root distribution, water and nutrient uptake are estimated and not dynamic in most TBMs. Root traits can provide a framework for scaling dynamic root functions (such as fine root proliferation, loss of root conductivity, or hydraulic redistribution) into TBMs to improve model veracity – a pathway indicated by the large arrow.

340x290mm (150 x 150 DPI)

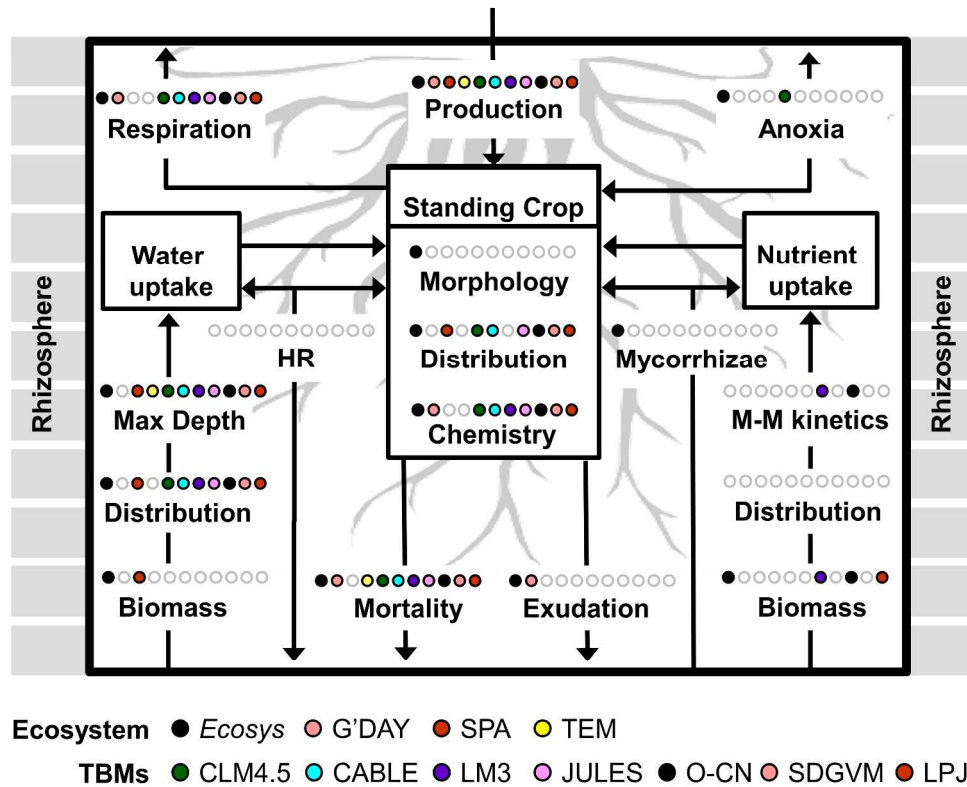


Figure 4. Key root structural and functional attributes and their inclusion in several well-known ecosystem and terrestrial biosphere models (TBMs) – filled circles represent model inclusion. Dynamic root functions such as Michaelis-Menten (M-M) nutrient uptake kinetics, hydraulic redistribution of water (HR) and downregulation due to low oxygen (Anoxia) are rarely included in the models. Other functions such as water uptake are widely represented when linked specifically to root depth, but rarely consider actual root biomass. Model references as in Tables 2, 3.

963x861mm (81 x 81 DPI)

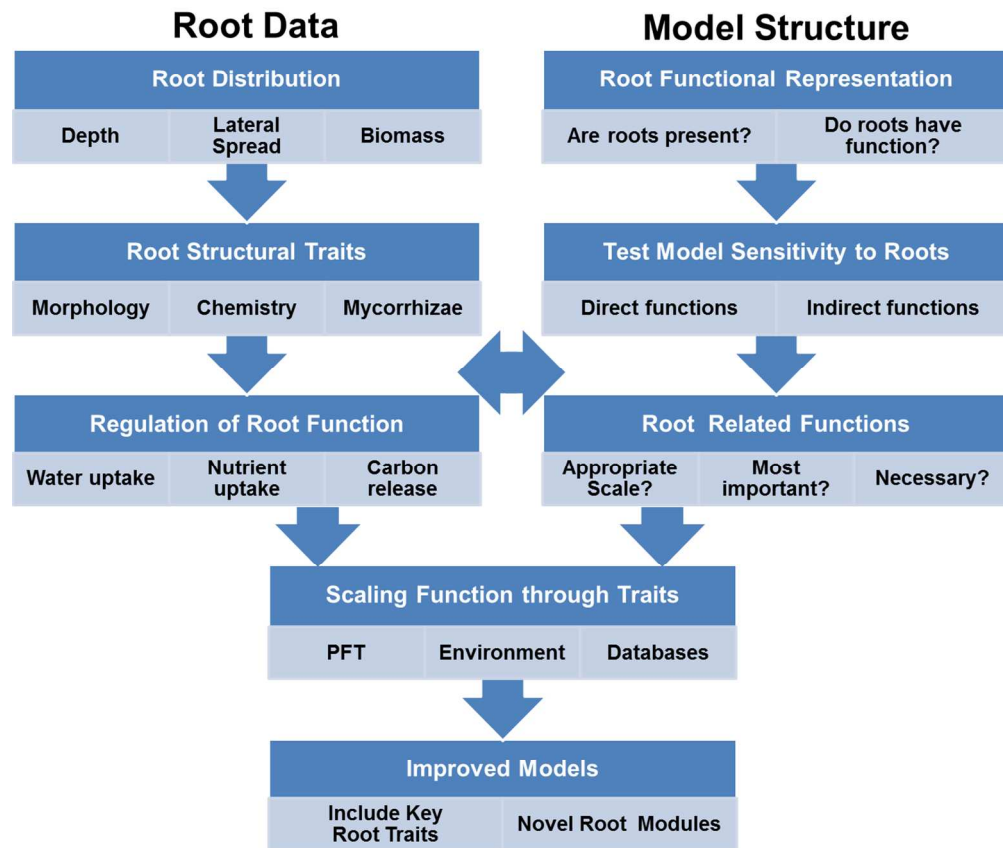


Figure 5. Framework for assessment of root data, and its importance in scaling ecosystem function through root traits for modeling the terrestrial biosphere. (left) Root distribution is the most common dataset available, and is used in many TBMs to regulate water use (Fig 4). Improved modeling will include root structural traits (e.g., size, age, order, display, C:N, mycorrhizal associations), and their associated functions (e.g., water and nutrient uptake, and carbon release through respiration, exudation and turnover). (right) Model evaluation should first assess the presence of roots or root functions, including both direct (e.g., water uptake based on root distribution) and indirect (e.g., nitrogen uptake based on plant demand) functions. Efforts must be made to understand the role of roots for specific processes at the appropriate spatial and temporal scales (Fig 3). Key root functions should be prioritized based on current mechanistic knowledge of root processes and dynamic biotic/abiotic regulation of those processes, as well as by their relative importance to the model. Addition of new root functionality to a model will require development of trait databases that can be scaled across landscapes based on species and plant functional type (PFT) characteristics, soil and environmental conditions.

239x210mm (150 x 150 DPI)